



Iris
Biotech



COUPLING REAGENTS



Version: IB12_1

Empowering Peptide Innovation

With this guiding theme in mind, Iris Biotech's mission is to support researchers by supplying

- innovative technologies,
- rare compounds,
- as well as a broad portfolio on standard consumables,

available in flexible quantities from small scale to bulk quantities. To fulfill our dedication "Empowering Peptide Innovation", we are attending various conferences, symposia, and exhibitions each year. This allows us to remain in direct contact with scientists all over the world, both from academia and industry, to exchange knowledge, and to gather new ideas to tackle your current challenges.

Guided by our dedication to provide

- competent service,
- as well as novel substances and
- latest technologies,

Iris Biotech is your trusted partner for the world of peptides, while having strong expertise in associated disciplines. Thus, our portfolio comprises reagents and tools for the synthesis and modification of peptides, e.g., amino acids, resins and solvents but also for related technologies such as drug delivery, linkerology® and life sciences.

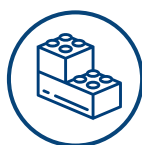
Owed to the growing demand for tailor-made compounds, our portfolio is fine-tuned by our custom synthesis service at Iris Biotech Laboratories. Our skilled scientists offer profound expertise in

- *de novo* route development,
- upscaling towards larger scale production,
- as well as synthesis optimization for increased efficiency.

Examples are the synthesis of rare chiral building blocks, unnatural amino acid derivatives, sophisticated orthogonal protecting groups, heterocycles, building blocks for nucleotides, PEGs and PEG-analogs as well as specific linkers for controlled drug delivery and release.



Amino Acids



Building Blocks



Life Sciences



Drug Delivery



Reagents



Resins



Linkerology®



Click Chemistry

Portfolio Overview

Peptide Synthesis and Modification

(Protected) Amino Acids

Standards such as Fmoc-D/L-AAA and Boc-D/L-AAA, Smoc amino acids for peptide synthesis in water, variety of protecting groups (e.g., Pbf, Trt, ^tBu, Bzl, Acn, Mob, SIT, Phacn, Allocam, Mmt), unusual amino acids, fluorinated derivatives, substituted prolines, arginine analogs

Building Blocks

Amino alcohols, amino aldehydes, diamines and hydrazines, (pseudoproline) dipeptides, polyamines and spermines, fatty acid derivatives, peptide nucleic acids (PNAs)

Reagents

Coupling reagents, solvents and scavengers, protecting groups

Resins

Preloaded resins (e.g., based on Trityl, TCP, TentaGel, Methoxybenzhydryl, Merrifield, PAM, Rink, Wang), scavenger resins, hydrazone resins, poly(acrylamide) resins, Cyclover

Linkerology® and Drug Delivery

Linkers for Solid Phase Peptide Synthesis

Cleavable Linkers

Val-Ala-based, Val-Cit-based, disulfide-based, Dde-helping hands, pH-sensitive linkers

Photo-Activatable Linkers

Functionalized Linkers

Clickable linkers, trifunctional linkers, linkers with maleimide function, cross-linkers, selective N-term acylation and biotinylation, 5HP2O

PROTACs

Ligands, linkers & modules

Fullerenes, Poly(2-oxazolines), Dextrans & Plant-Derived Cholesterol

Superparamagnetic Iron Oxide Nanoparticles

Poly-Amino Acids

Poly-Arg, Poly-Glu, Poly-Lys, Poly-Orn, Poly-Sar

PEGylation

Branched PEGylating reagents, (amino-)PEG-acids, PEG-amines & hydrazides & guanidines, reagents for Click-conjugation, Biotin-PEG-reagents, PEG-thiols, PEG-maleimides, other PEGylating reagents

Life Sciences

Biotinylation Reagents

Carbohydrates

Galactose, Glucose, Mannose, Xylose and others

Drug Metabolites

Peptides

Substrates & Inhibitors

E.g., protein kinase inhibitors, substrates for fusion (Halo/Snap/Clip)-tagged proteins

Natural Products

Dyes and Fluorescent Labels

E.g., ICG, AMC, DAPI

Maillard & Amadori Reaction Products

Large portfolio of derivatives useful as standards for food, pharma and cosmetics industry

Vitamins

Custom Synthesis

Your project requires a compound not listed in our portfolio?
Get in contact and inquire about our custom synthesis capabilities.

Our experienced scientists are excited to accept your synthetic challenge!

In such cases, your request undergoes the following stages:



Step-by-Step Analysis

- Customer's demands

Process Evaluation

- Detailed literature review
- Synthetic possibilities

Strategy Development

- Protocol development
- Method development and validation
- Customized synthesis

Quality Consistency

- Identity confirmation
- Purity verification

Our Service Promise

All our services are based on high standards, transparency & documentation, trust, honesty & confidentiality, as well as the required know-how.

High Standards

- Values: sustainability & responsibility
- State-of-the-art equipment & latest technologies
- High quality standards
- Qualified suppliers & regular audits

Transparency & Documentation

- Talk to our specialists – customer care
- Certificates of analysis & origin
- Impurity profiling
- Safety data sheets
- Analytical and process reports

Trust, Honesty & Confidentiality

- Intergenerational business valuing partnerships
- Meeting the customer's expectations
- Integrity towards our customers

Our Know-How

- One-step reactions & complex multi-step synthesis
- Scalability from mg to kg quantities
- Route scouting



Table of Contents

1. Introduction	1
------------------------	----------

2. Carbodiimides	3
-------------------------	----------

3. Phosphonium Salts	11
-----------------------------	-----------

4. Uronium and Aminium Salts	16
-------------------------------------	-----------

5. Acid-Halides	23
------------------------	-----------

6. Anhydrides	28
----------------------	-----------

7. Triazines	30
---------------------	-----------

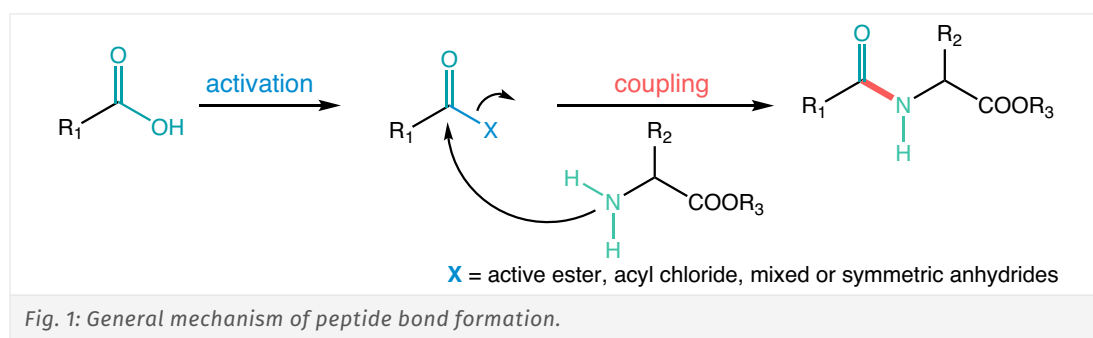
8. Ynamides	33
--------------------	-----------

Index	36
Code of Conduct	39
Terms and Conditions of Sales	41

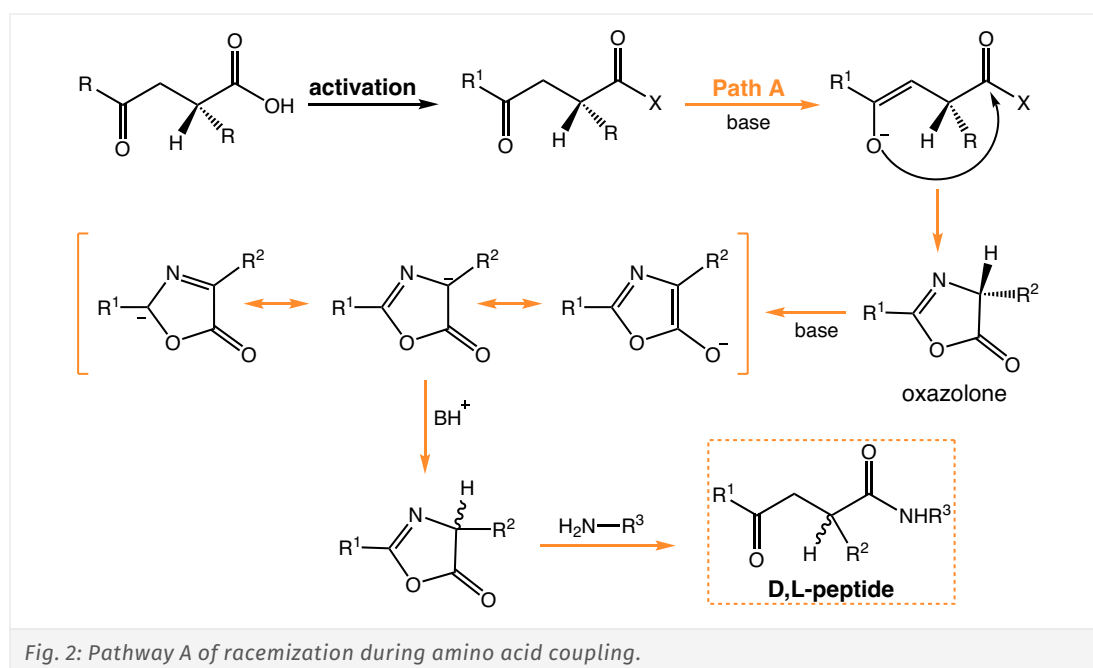
1. Introduction

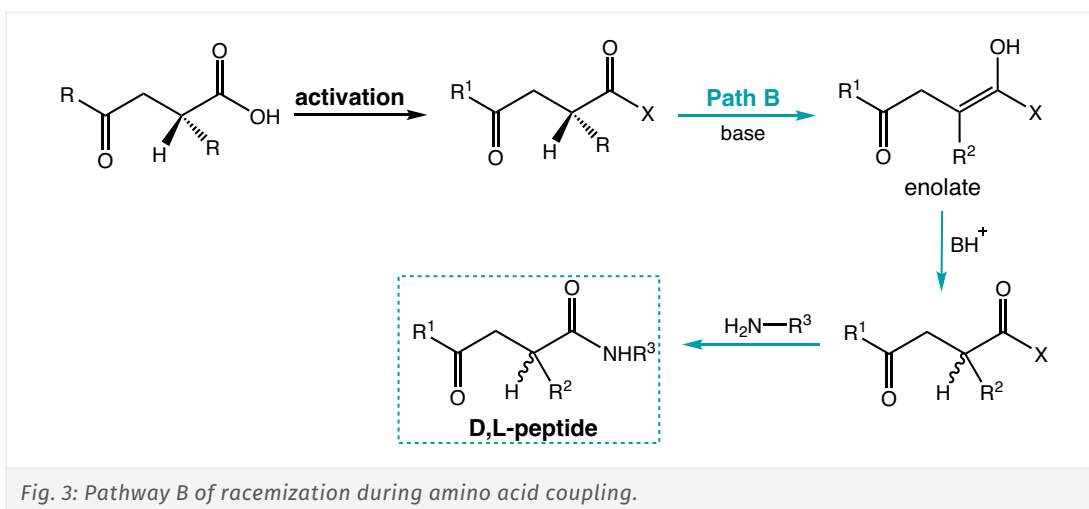
The naturally low reactivity of functional groups in amino acids makes direct coupling inefficient. To address this, coupling reagents are employed to activate the carboxyl group of one amino acid, enhancing its electrophilic character and promoting nucleophilic attack by the amino group of another.

The primary function of a coupling reagent is to transform the carboxyl group into a more reactive intermediate, typically by incorporating an electron-withdrawing group. This activated species then undergoes a nucleophilic substitution with the free amino group of the incoming amino acid (aminolysis), forming a peptide bond (amide bond) and releasing a by-product. For successful peptide bond formation, the coupling reagent must be highly reactive and selective, minimize undesirable side reactions, and generate soluble by-products that can be easily removed from the reaction mixture.

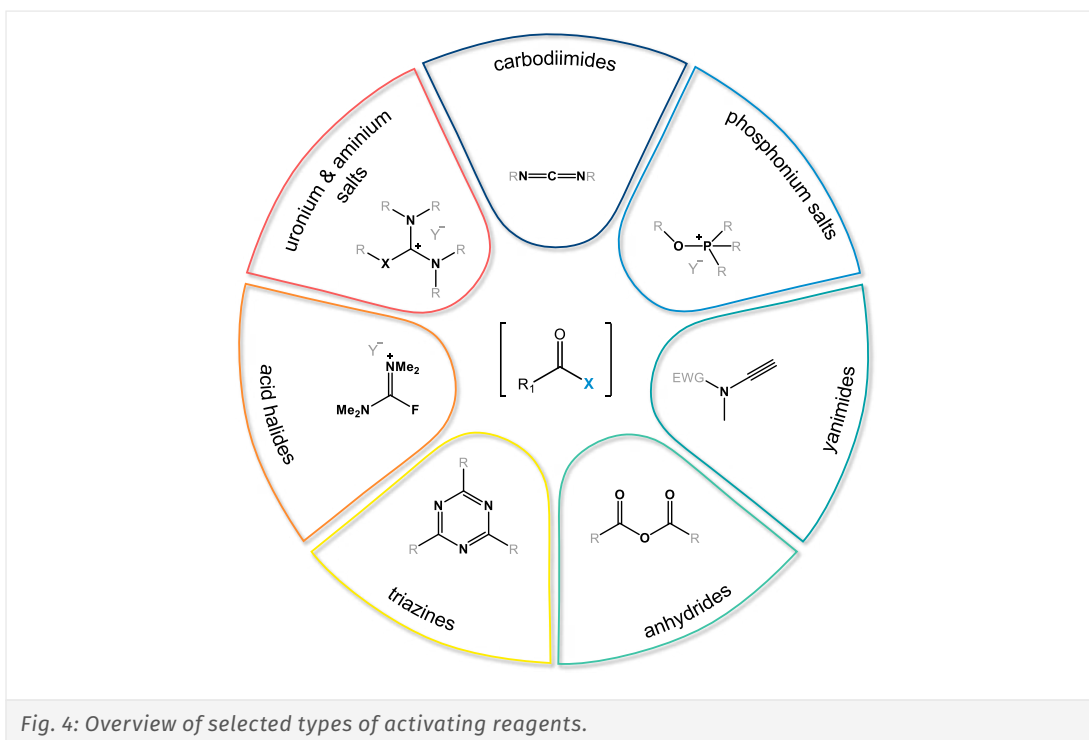


However, the activation step poses a significant challenge, as there is a risk of racemization, which can proceed via two possible pathways (see Fig. 2, Fig. 3): First, through an intramolecular cyclization reaction that leads to the formation of an oxazolone intermediate (path A, Fig. 2). Second, through direct enolization (path B, Fig. 3).





This racemization risk is critical in peptide synthesis, as both pathways – cyclization and enolization – can result in the loss of the desired chiral integrity. A careful consideration of reaction conditions is therefore essential. For this purpose, several coupling reagents have been developed, each with unique properties and mechanisms. The most commonly used coupling reagents can be classified into different categories (see Fig. 4).



References:

- Peptide Coupling Reagents, More than a Letter Soup; A. El-Faham, F. Albericio; **Chem. Rev.** 2011; **111**: 6557-6602. <https://doi.org/10.1021/cr100048w>
- Recent development in peptide coupling reagents; T. I. Al-Warhi, H. M. A. Al-Hazimi, A. El-Faham; **Journal of Saudi Chemical Society** 2012; **16(2)**: 97-116. <https://doi.org/10.1016/j.jscs.2010.12.006>

[↑ back to content](#)

2. Carbodiimides

In this reaction, a carbodiimide reacts with a carboxylic acid to form *O*-acylurea, a highly reactive intermediate that enables aminolysis (peptide bond formation). In the presence of an excess of the carboxylic acid, the formed *O*-acylurea may also convert into a symmetrical anhydride. These compounds further facilitate peptide bond formation but at a slower rate. *O*-acylisourea has also been demonstrated to undergo cyclization, resulting in the formation of oxazolone, a species that exhibits reduced reactivity and increased propensity for racemization, both of which could potentially compromise the stereochemical integrity of the desired product (see Fig. 6).

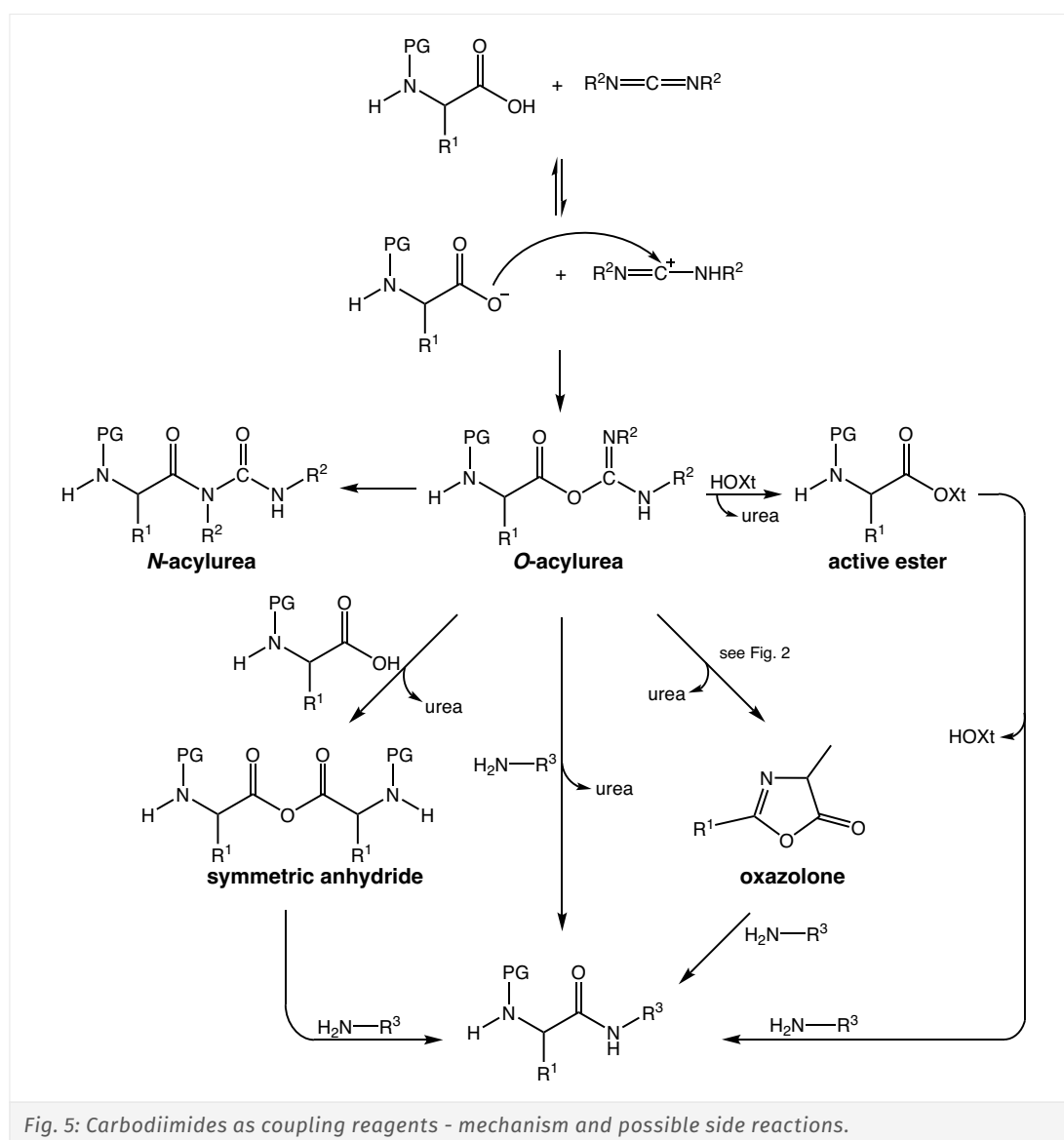


Fig. 5: Carbodiimides as coupling reagents - mechanism and possible side reactions.

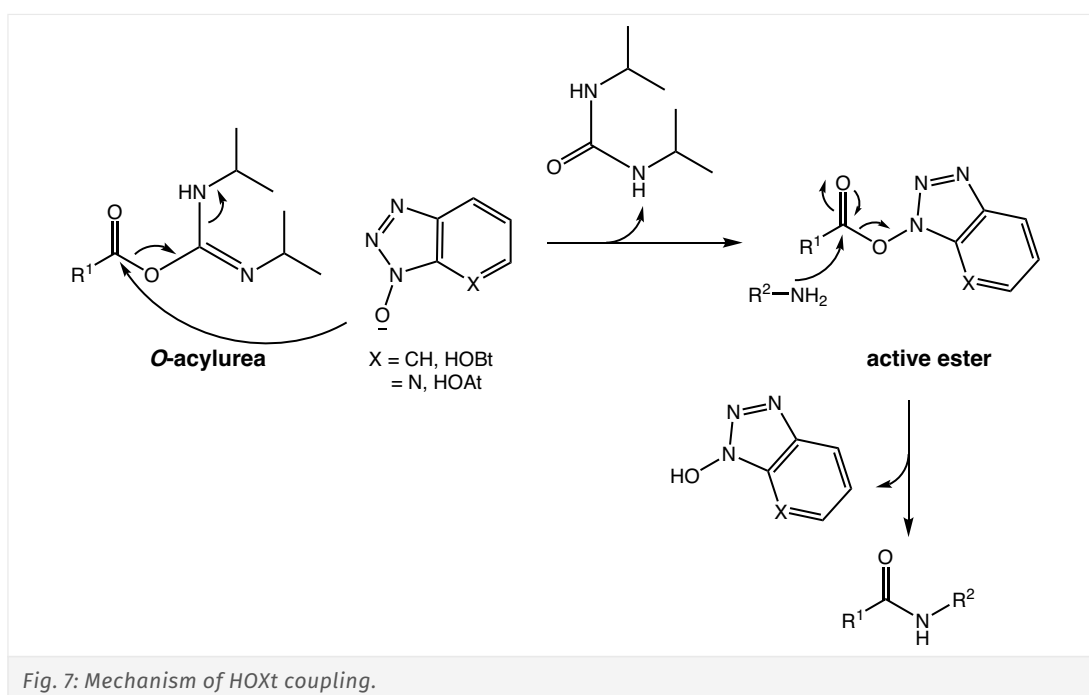
The carbodiimide method by applying reagents such as the symmetrical dicyclohexylcarbodiimide (DCC, [RL-1013 on page 9](#)) and diisopropylcarbodiimide (DIC, [RL-1015 on page 9](#)) or the unsymmetrical reagents *N*-ethyl-*N*'-(3-dimethylaminopropyl)carbodiimide (EDC, [RL-1022 on page 9](#)) or 1-*tert*-butyl-3-ethylcarbodiimide (TBEC, [RL-3910 on page 9](#)) remains one of the most widely used strategies for peptide bond formation.

A significant limitation of using DCC in carbodiimide-mediated couplings is the formation of *N*-acylurea, an unreactive by-product that consumes valuable starting materials without contributing to the peptide yield, e.g., in polar solvents such as DMF. Furthermore, DCC generates dicyclohexylurea (DCU), an insoluble side product that poses challenges for removal, even with chromatographic purification techniques.

These drawbacks have driven the development of alternative carbodiimides, such as diisopropylcarbodiimide (DIC, [RL-1015 on page 9](#)) which offer an improved solubility in organic solvents.

N-ethyl-*N'*-(3-dimethylaminopropyl)carbodiimide (EDC, [RL-1022 on page 9](#)) is especially advantageous in solution-phase reactions due to its water-soluble urea by-product which facilitates easier post-reaction workup and simplifies purification, e.g., by extraction methods.

To address challenges with side product formation, racemization and purification, carbodiimides are often used in combination with additives like 1-hydroxybenzotriazole (HOBt) or 1-hydroxy-7-azabenzotriazole (HOAt). These additives form more stable intermediates (active esters) which suppress the formation of *N*-acylurea and reduce racemization (see [Fig. 7](#)). The enhanced effectiveness of HOBt and HOAt is largely due to their ability to protonate the *O*-acylisourea intermediate, preventing unwanted intramolecular cyclization by forming the mentioned active esters. This shift in the reaction pathway helps to lower the risk of racemization in many cases.



HOAt offers different advantages over other additives due to the presence of a nitrogen atom at the 7-position of the benzotriazole ring (see [Fig. 8](#)). This nitrogen atom enhances the ester reactivity through two key mechanisms: first, its electron-withdrawing effect increases the overall reactivity of the intermediate; second, the strategic placement of the nitrogen at position 7 enables a classic neighboring group effect, further accelerating the reaction while preserving the configurational integrity of the peptide product. As a result, HOAt yields higher coupling efficiencies and significantly reduces racemization compared to other additives.

[↑ back to content](#)

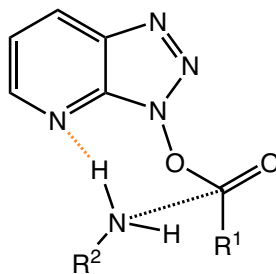


Fig. 8: Neighboring group effect of HOAt.

However, both HOBt and HOAt present safety concerns due to their low molecular weight and the presence of multiple consecutive nitrogen atoms that make them highly explosive. To address this issue, OxymaPure® has been developed as a safer alternative. OxymaPure® ([RL-1180 on page 9](#)) reduces racemization and side reactions while maintaining a high efficiency in peptide coupling protocols. It forms a stable active ester (see [Fig. 9](#)) and has been shown to outperform traditional additives like HOBt in many cases, offering a safer and more reliable option for peptide synthesis.

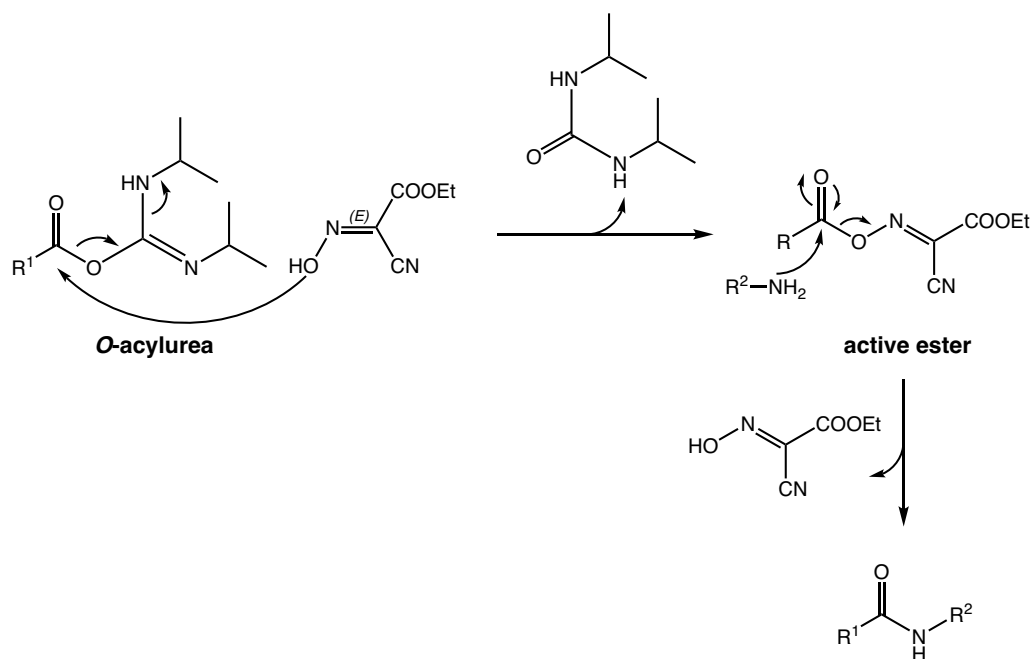
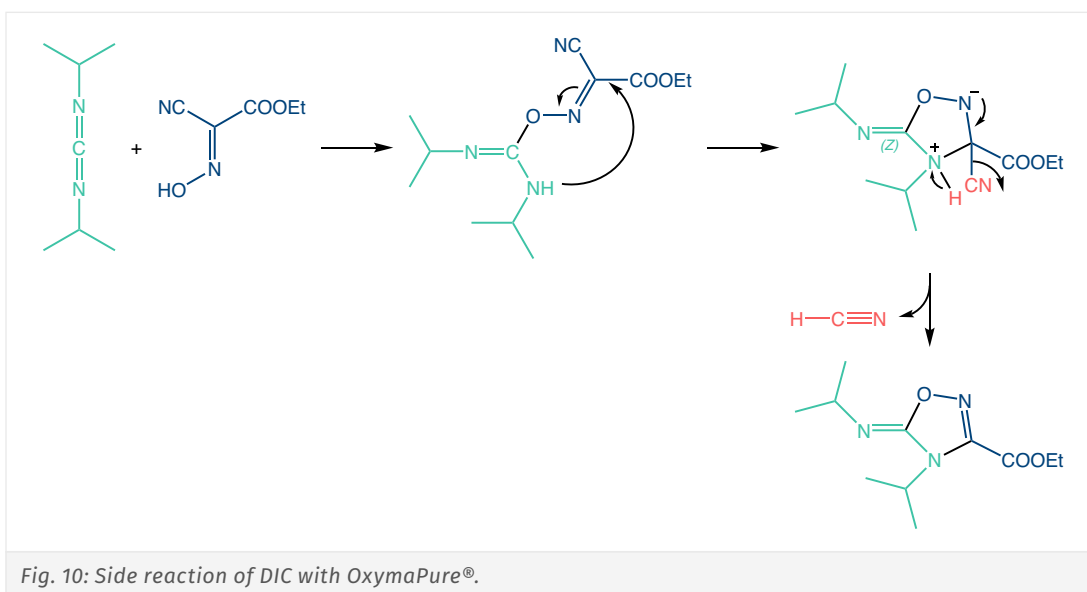
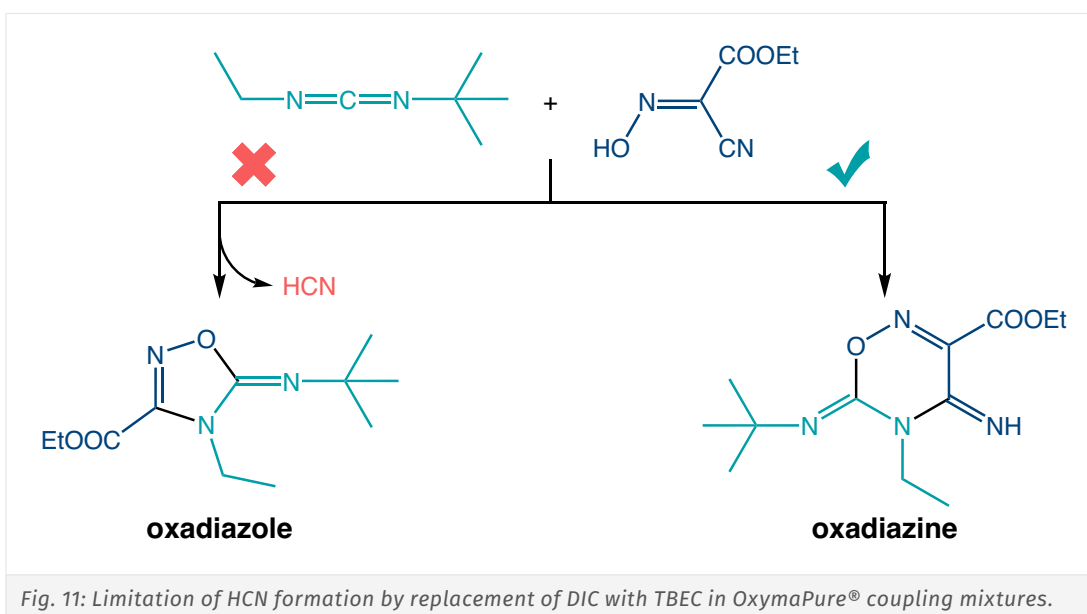


Fig. 9: Coupling mechanism with OxymaPure®.

Despite these benefits, a side reaction has been previously reported where OxymaPure® and DIC form a cyclic adduct, an oxadiazole, resulting in the release of hydrogen cyanide (HCN) as a by-product (see [Fig. 10](#)).



This side reaction presents potential safety concerns, but it occurs only to a limited extent. Moreover, new research has demonstrated that the use of asymmetric carbodiimides, such as 1-*tert*-Butyl-3-ethyl-carbodiimide (TBEC, [RL-3910 on page 9](#)) or EDC ([RL-1022 on page 9](#)), can effectively suppress the formation of HCN. In this case, instead of a 5-membered oxadiazole ring, a 6-membered oxadiazine ring is formed, reducing the likelihood of HCN release (see [Fig. 11](#)).



Oxyma-B ([RL-2290 on page 9](#)) is described as an alternative to HOAt, which, compared to OxymaPure®, can reduce racemization, for example in histidine, from 3% to 1%. For large-scale applications of Oxyma-B ([RL-2290 on page 9](#)), studies on potential side reactions have been conducted. Orlandin and colleagues demonstrated that a carbodiimide-mediated Beckmann rearrangement might occur. This side reaction might lead to a capping of the peptide and therefore limit the yield of the desired peptide. Additionally, this side reaction might cause truncated peptide sequences.

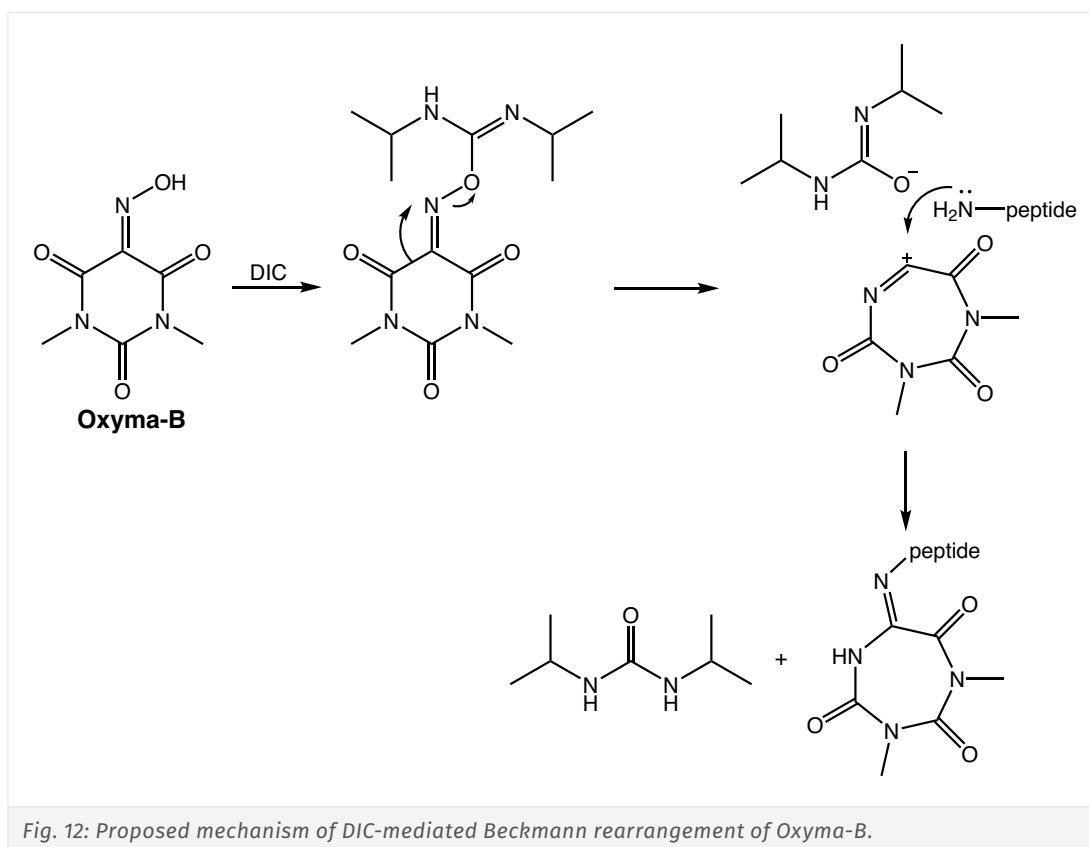


Fig. 12: Proposed mechanism of DIC-mediated Beckmann rearrangement of Oxyma-B.

Guidelines for standard carbodiimide coupling conditions at room temperature

Carbodiimide + additive method

- I. In a separate flask or tube, dissolve the protected amino acid (5 eq. relative to the resin loading) and HOxt (5 eq.) in DMF or NMP (5 mL/g of resin).
- II. Add carbodiimide (5 eq.) to the solution and stir the mixture for 10 to 20 min for pre-activation.
- III. Transfer the activated coupling mixture to the swollen resin and shake the mixture according to the following guidelines:

Conditions	Reaction time [min]
Standard	45
Difficult coupling (e.g., bulky side-chain)	90
After incorporation of the first 20 aa	2 x 45
After sterically-hindered aa	2 x 45

- V. Remove the excess reagents after the desired reaction time.
- VI. Wash resin with DMF (at least 3 x 5 mL).

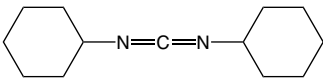

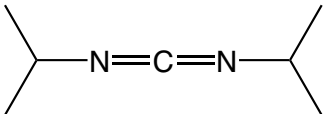

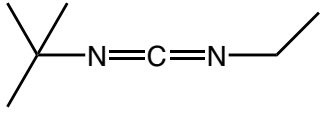

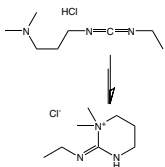

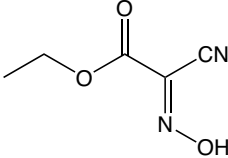

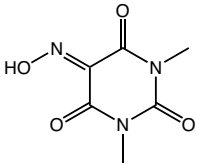

Note:

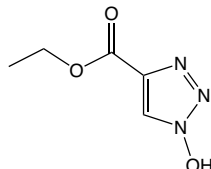

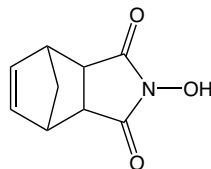

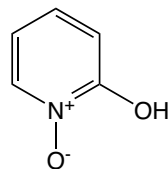

- The active ester formed is unstable and should not be stored; it is recommended to prepare the reaction mixture fresh for each step.
- OxymaPure® (RL-1180 on page 9) has been reported to outperform HOBt, though not consistently HOAt. It can also be used with DIC without requiring pre-activation, to avoid HCN formation. As mentioned earlier TBEC (RL-3910 on page 9) can serve as a replacement for DIC (RL-1015 on page 9). This combination may also eliminate the need for pre-activation in certain cases.
- Coupling protocols for the DIC (RL-1015 on page 9) /OxymaPure® (RL-1180 on page 9) method using microwave heating are available, and it is advisable to follow the manufacturer's guidelines for optimal results.

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- Amide Formation: Choosing the Safer Carbodiimide in Combination with Oxyma Pure to Avoid HCN Release; S. R. Manne, O. Luna, G. A. Acosta, M. Royo, A. El-Faham, G. Orosz, B. G. de la Torre, F. Albericio; **Org. Lett.** 2021; **23(17)**: 6900-6904. <https://doi.org/10.1021/acs.orglett.1c02466>
- Oxyma-B, an excellent racemization suppressor for peptide synthesis; Y. E. Jad, S. N. Khattab, B. G. de la Torre, T. Govender, H. G. Kruger, A. El-Faham, F. Albericio; **Org. Biomol. Chem.** 2014; **12**: 8379-8385. <https://doi.org/10.1039/C4OB01612B>
- Carbodiimide-Mediated Beckmann Rearrangement of Oxyma-B as a Side Reaction in Peptide Synthesis; A. Orlandin, I. Guryanov, L. Ferrazzano, B. Biondi, F. Biscaglia, C. Storti, M. Rancan, F. Formaggio, A. Ricci, W. Cabri; **Molecules** 2022; **27(13)**: 4235. <https://doi.org/10.3390/molecules27134235>

Coupling Reagents

			Product details
RL-1013	DCC		
<i>N,N'</i> -Dicyclohexylcarbodiimide			
CAS-No.	538-75-0		
Formula	C ₁₃ H ₂₂ N ₂		
Mol. weight	206,3 g/mol		
			
RL-1015	DIC		
<i>N,N'</i> -Diisopropylcarbodiimide			
CAS-No.	693-13-0		
Formula	C ₇ H ₁₄ N ₂		
Mol. weight	126,20 g/mol		
			
RL-3910	T-BEC		
1- <i>tert</i> -Butyl-3-ethylcarbodiimide			
CAS-No.	1433-27-8		
Formula	C ₇ H ₁₄ N ₂		
Mol. weight	126,20 g/mol		
			
RL-1022	EDC*HCl		
<i>N</i> -Ethyl- <i>N'</i> -(3-dimethylaminopropyl)carbodiimide hydrochloride			
CAS-No.	25952-53-8		
Formula	C ₈ H ₁₇ N ₃ *HCl		
Mol. weight	155,24*36,45 g/mol		
			
RL-1180	OxymaPure		
Ethyl cyano(hydroxyimino)acetate, Ethyl cyano-glyoxylate-2-oxime			
CAS-No.	3849-21-6		
Formula	C ₅ H ₆ N ₂ O ₃		
Mol. weight	142,11 g/mol		
			
RL-2290	Oxyma-B		
5-(Hydroxyimino)1,3-dimethylpyrimidine-2,4,6-(1H,3H,5H)-trione			
CAS-No.	5417-13-0		
Formula	C ₆ H ₇ N ₃ O ₄		
Mol. weight	185,14 g/mol		
			

			Product details
RL-1091	HOCT		
Ethyl 1-hydroxy-1H-1,2,3-Triazole-4-carboxylate			
CAS-No.	137156-41-3		
Formula	C ₅ H ₇ N ₃ O ₃		
Mol. weight	157,13 g/mol		
RL-1036	HONB		
N-Hydroxy-5-norbornene-2,3-dicarboxylimide			
CAS-No.	21715-90-2		
Formula	C ₉ H ₉ NO ₃		
Mol. weight	179,17 g/mol		
RL-2002	HOPO		
2-Hydroxypyridine-N-oxide			
CAS-No.	13161-30-3		
Formula	C ₅ H ₅ NO ₂		
Mol. weight	111,10 g/mol		


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Introduction

Carbodimides

Phosphonium Salts

Uronium and Aminium Salts

Acid-Halides

Anhydrides

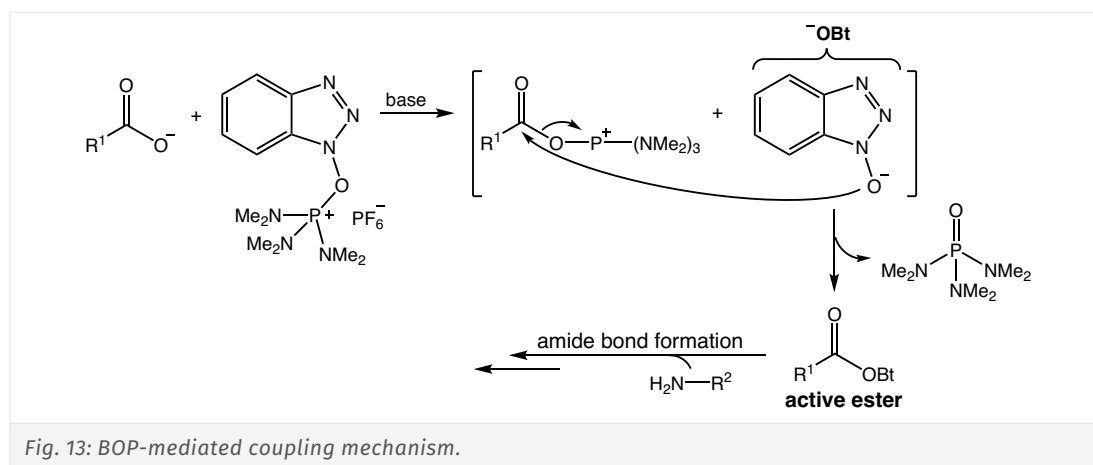
Triazines

Ynamides

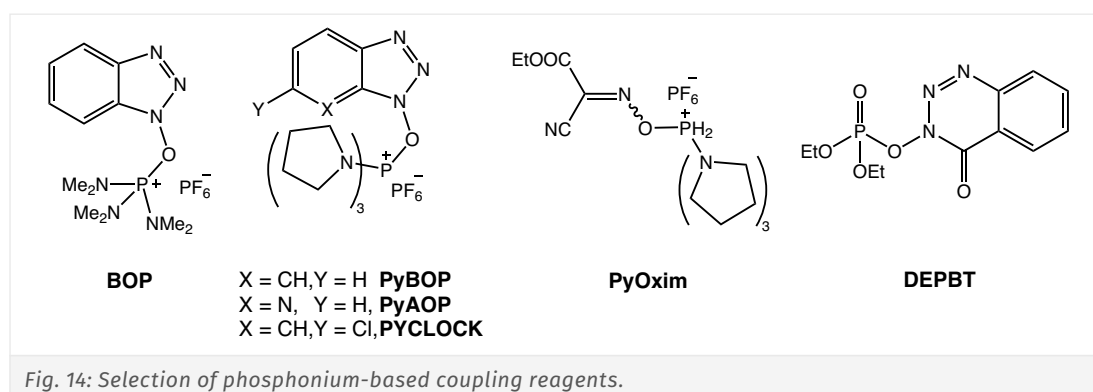
Index

3. Phosphonium Salts

Phosphonium salts activate the carboxyl group of the amino acid by forming a phosphonium or benzo-triazolyl ester as the activated intermediate. This compound in turn reacts with the amine group of the growing peptide chain to form an amide bond, with minimal side reactions. These couplings are performed in the presence of at least 1 equivalent (eq.) of base (most commonly 2 eq. of diisopropylethylamine (DIPEA) or *N*-methylmorpholine (NMM) are used), as the phosphonium-based reagents react with a carboxylate. It is noteworthy that unlike carbodiimide-based couplings, phosphonium salts avoid urea formation thereby reducing the likelihood of side product generation.

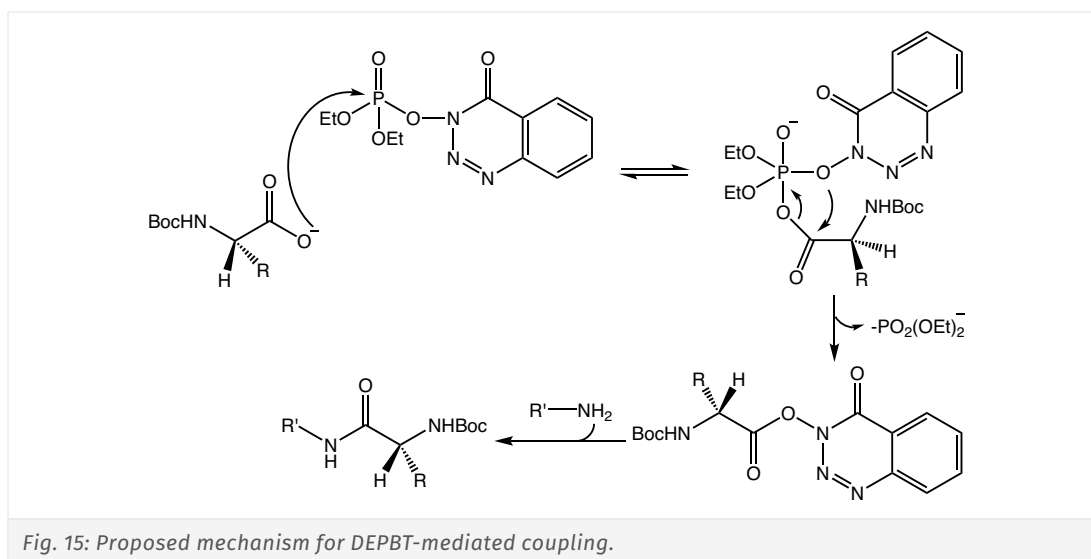


Benzotriazol-1-yloxytris(dimethylamino)phosphonium hexafluorophosphate (BOP, [RL-5096 on page 14](#)) was the first phosphonium salt introduced for peptide synthesis. However, its use raised safety concerns due to the formation of hexamethylphosphoramide (HMPA), a carcinogenic side product (see [Fig. 13](#)). As a result, safer alternatives like benzotriazol-1-yloxytri(pyrrolidino) phosphonium hexafluorophosphate (PyBOP, [RL-2005 on page 15](#)) and [(7-azabenzotriazol-1-yl)oxy]tris(pyrrolidino) phosphonium hexafluorophosphate (PyAOP, [RL-1210 on page 15](#)) were developed in which the dimethylamine moiety was replaced by pyrrolidine to avoid HMPA formation. These newer reagents gained popularity due to their lower risk of racemization in comparison to the carbodiimide method and high efficiency in challenging peptide couplings. Bromotri(pyrrolidino) phosphonium hexafluorophosphate (PyBrOP, [RL-1051 on page 15](#)) and (6-chloro-benzotriazol-1-yloxy)tris(pyrrolidino) phosphonium hexafluorophosphate (PYCLOCK, [RL-2180 on page 15](#)) are described in the coupling of *N*-methylated amino acids.



After benzotriazoles like HOBt and HOAt were classified as explosive, and OxymaPure® ([RL-1180 on page 9](#)) was introduced as a safer alternative, *O*-[(cyano(ethoxycarbonyl)methyliden)-amino]-xyloxytri(pyrrolidino) phosphonium hexafluorophosphate (PyOxim, [RL-2270 on page 14](#)), an OxymaPure®-based phosphonium salt was developed. This new reagent was designed to offer improved safety while maintaining high efficiency in peptide coupling reactions.

3-(Diethoxyphosphoryloxy)-1,2,3-benzotriazin-4(3H)-one (DEPBT, [RL-1153 on page 14](#)) is an organo-phosphorus compound that is described as a coupling reagent in the synthesis of bioactive peptides with a great resistance to racemization.



Guidelines for standard phosphonium-based coupling conditions

BOP ([RL-5096 on page 14](#))

- I. In a separate flask or tube, dissolve the *N*-protected amino acid (2 eq. relative to the resin loading) in DMF (5 mL/g of resin) and add it to the pre-swelled resin.
- II. Add BOP (2 eq., 1 M solution in DMF) and DIPEA or NMM (4 eq.) to the solution. Note: In order to suppress racemization HOBt or OxymaPure® (2 eq., 0.5 M solution in DMF) can be added.
- III. Shake the resin with the coupling mixture for 10-60 min.
- IV. Remove the excess reagents after the desired reaction time.
- V. Wash the resin with DMF (at least 3 x 5 mL).

PyBOP ([RL-2005 on page 15](#))

- I. In a separate flask or tube, dissolve the *N*-protected amino acid (1.1 eq. relative to the resin loading) in DMF (5 mL/g of resin) and add it to the pre-swelled resin.
- II. Add PyBOP (1.1 eq., 1 M solution in DMF) and DIPEA or NMM (2 eq.) to the solution. Note: In order to suppress racemization HOBt or OxymaPure® (1.1 eq., 0.5 M solution in DMF) can be added.
- III. Shake the resin with the coupling mixture for 30-60 min.
- IV. Remove the excess reagents after the desired reaction time.
- V. Wash the resin with DMF (at least 3 x 5 mL).

[↑ back to content](#)

PyBrOP (RL-1051 on page 15) or PyCLOCK (RL-2180 on page 15) for *N*-methylated amino acids

- I. In a separate flask or tube, dissolve the *N*-protected amino acid (2 eq. relative to the resin loading) in DMF or DCM (5 mL/g of resin) and add it to the resin.
- II. Add PyBrOP or PyCLOCK (1.1 eq., 1 M solution in DMF) and cool the solution to 0°C.
- III. Add DIPEA (6 eq.) to the reaction mixture.
- IV. Shake the resulting reaction mixture for 1 min at 0°C followed by 60 min at room temperature.
- V. Remove the excess reagents after the desired reaction time.
- VI. Wash the resin with DMF (at least 3 x 5 mL).

PyOxim (RL-2270 on page 14)

- I. In a separate flask or tube, dissolve the *N*-protected amino acid (3 eq. relative to the resin loading) in DMF or NMP (5 mL/g of resin).
- II. Add PyOxim (3 eq.) and DIPEA (6 eq.) to the solution.
- III. Shake the resulting reaction mixture for 1-2 min (pre-activation).
- IV. Add the solution to the resin and allow the coupling to proceed for 30 min.
- V. Remove the excess reagents after the desired reaction time.
- VI. Wash the resin with DMF (at least 3 x 5 mL).

DEPBT (RL-1153 on page 14) (solid phase)

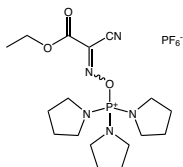

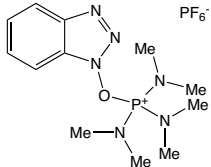

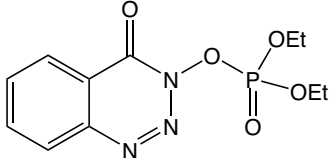

- I. In a separate flask or tube, dissolve the *N*-protected amino acid (1.5 eq. relative to the resin loading) in DCM or DMF (5 mL/g resin) and add it to the pre-swelled resin.
- II. Add DIPEA or NEt₃ (3 eq.).
- III. Add DEPBT (1.5 eq.).
- IV. Shake the resulting reaction mixture for 1 to 2 h.
- V. Remove the excess reagents after the desired reaction time.
- VI. Wash the resin with DMF or DCM (at least 3 x 5 mL).


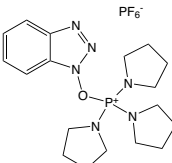

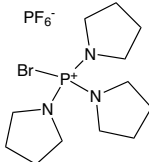

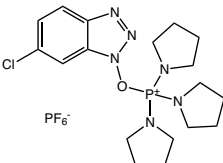

DEPBT (RL-1153 on page 14) (solution phase)

- I. Dissolve the protected amino acid and amino acid ester in THF or DMF (2 mL/mmol).
- II. Add DEPBT (1.1 eq.).
- III. Add NEt₃ (2 eq.).
- IV. Shake the resulting reaction mixture for 2 to 6 h.
- V. Pour the mixture into a saturated sodium chloride solution (40 mL/mmol) and extract the precipitate with ethyl acetate (3 x 50 mL).
- VI. In case of THF: triethylamine hydrochloride is removed by filtration.
- VII. Remove the solvent *in vacuo*.

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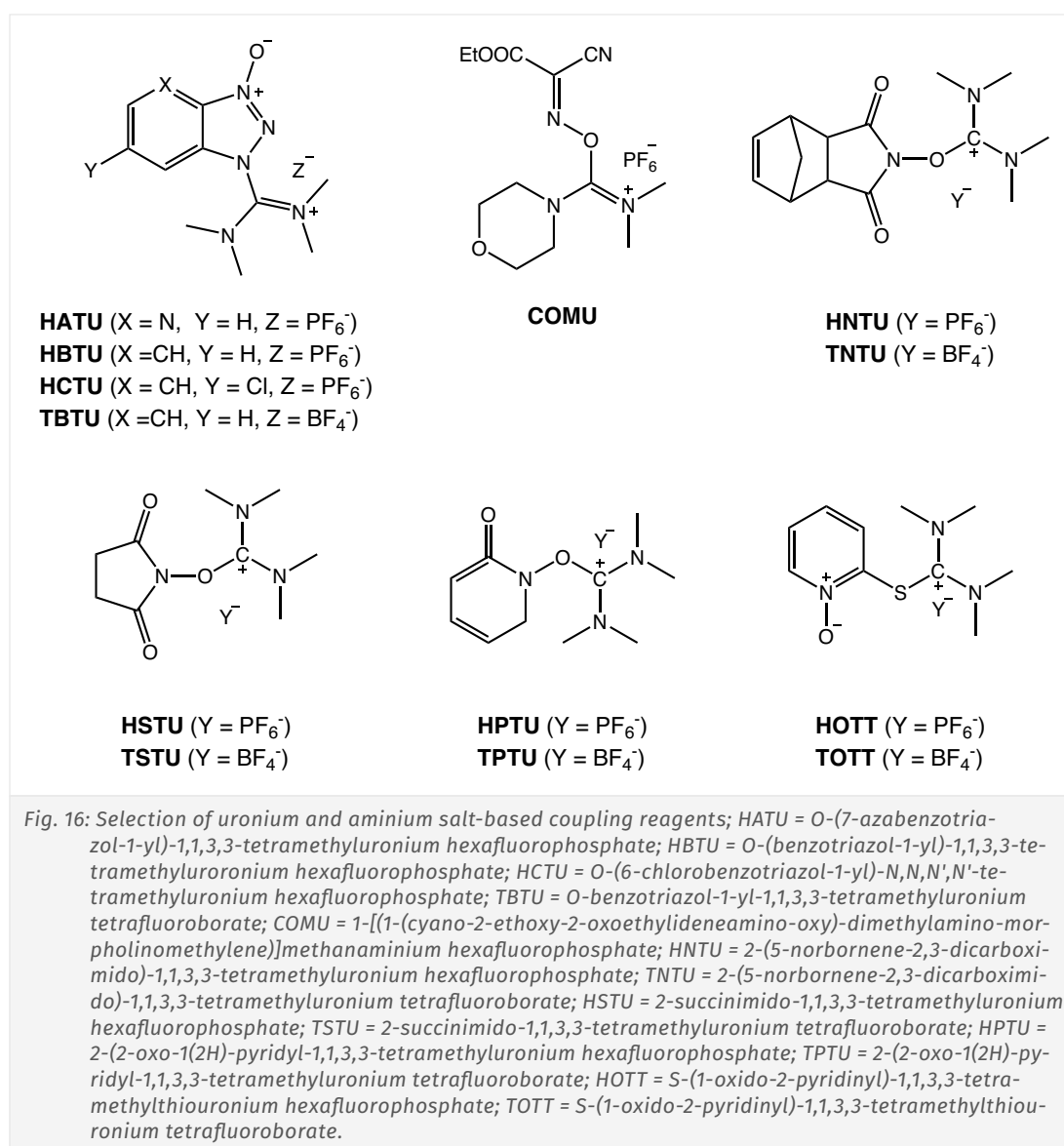
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		Product details
RL-2270 PyOxim [Ethyl cyano(hydroximino)acetato-O]tri-1-pyrrolidinyl-phosphonium hexafluorophosphate CAS-No. 153433-21-7 Formula C ₁₇ H ₂₉ N ₃ O ₃ P*PF ₆ Mol. weight 382,42*144,96 g/mol		
RL-5096 BOP Benzotriazol-1-yl-oxy-tris-(dimethylamino)-phosphonium hexafluorophosphate CAS-No. 56602-33-6 Formula C ₁₂ H ₂₂ N ₆ OP*PF ₆ Mol. weight 297,32*144,96 g/mol		
RL-1153 DEPBT 3-(Diethoxyphosphoryloxy)-1,2,3-benzotriazin-4(3H)-one CAS-No. 165534-43-0 Formula C ₁₁ H ₁₄ N ₃ O ₅ P Mol. weight 299,23 g/mol		

			Product details
RL-1210	PyAOP		
(3-Hydroxy-3H-1,2,3-triazolo[4,5-b]pyridinato-O) tri-1-pyrrolidinyl-phosphorus hexafluorophosphate			
CAS-No.	156311-83-0		
Formula	C ₁₇ H ₂₇ N ₇ OP*PF ₆		
Mol. weight	376,42*144,96 g/mol		
RL-2005	PyBOP		
Benzotriazole-1-yl-oxy-tris-pyrrolidino-phosphonium hexafluorophosphat			
CAS-No.	128625-52-5		
Formula	C ₁₈ H ₂₈ N ₆ OP*PF ₆		
Mol. weight	375,43*144,96 g/mol		
RL-1051	PyBrOP		
Bromo-tris-pyrrolidino-phosphonium hexafluoro-phosphat			
CAS-No.	132705-51-2		
Formula	C ₁₂ H ₂₄ N ₃ Br*PF ₆		
Mol. weight	321,22*144,96 g/mol		
RL-2180	PyCLOCK		
(6-Chlorobenzotriazol-1-yloxy)tripyrrolidinophosphonium hexafluorophosphate			
CAS-No.	893413-42-8		
Formula	C ₁₈ H ₂₇ ClN ₆ OP*PF ₆		
Mol. weight	409,87*144,96 g/mol		

4. Uronium and Aminium Salts

Uronium and aminium salts, commonly represented by *O*-(benzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate (HBTU, [RL-1030 on page 20](#)), *O*-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate (HATU, [RL-1190 on page 20](#)), *O*-benzotriazol-1-yl-1,1,3,3-tetramethyluronium tetrafluoroborate (TBTU, [RL-1060 on page 21](#)), and 1-[(1-(cyano-2-ethoxy-2-oxoethylideneamino-oxy)-dimethylamino-morpholinomethylene)]methanaminium hexafluorophosphate (COMU, [RL-1175 on page 20](#)), are derived from carbodiimides (see [Fig. 16](#)). Instead of a phosphonium residue they bear a positive charged carbon or nitrogen atom. Typically, tetrafluoroborate (BF_4^-) or hexafluorophosphate (PF_6^-) anions are used as non-nucleophilic counter ions. In comparative studies it was revealed that the counterion did not exert a significant influence on the rate of coupling or the rate of racemization. The first described compounds were HOBt derivatives analogous to the phosphonium-based coupling reagents.



Coupling Reagents

Like phosphonium salts uronium/aminium salts work in combination with a base such as DIPEA to activate the carboxyl function of the incoming amino acid (see Fig. 17).

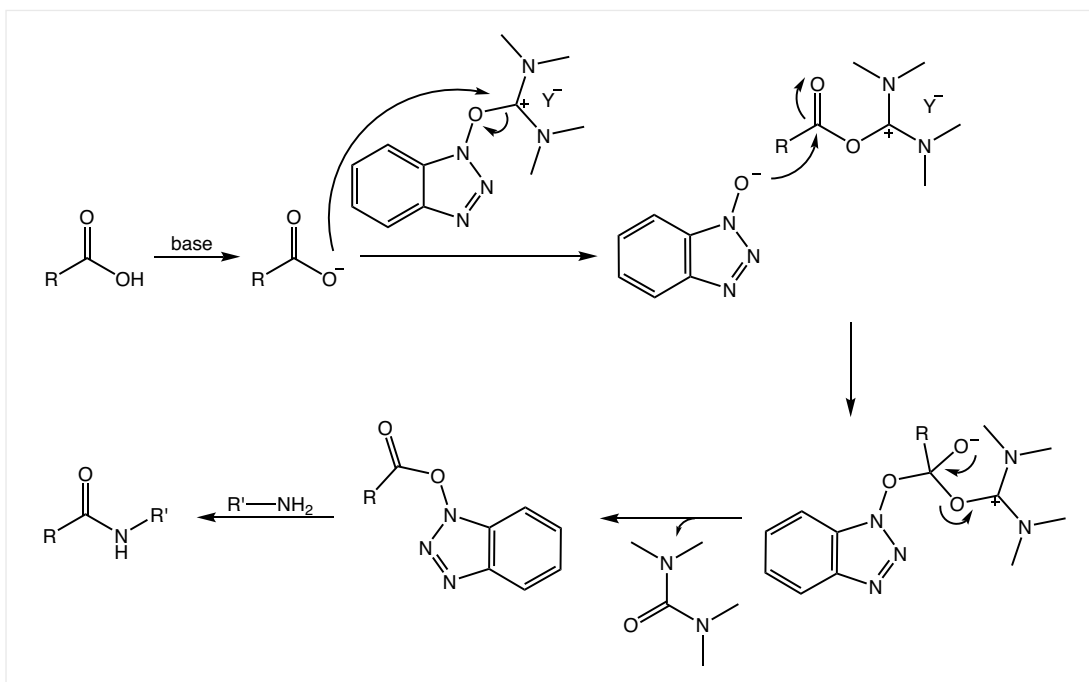


Fig. 17: Coupling mechanism of uronium reagents: The example of HBTU (RL-1030 on page 20).

However, these reagents might also directly react with the free amino function to form a guanidine by-product, which in turn leads to truncated peptide sequences and reduces the yield of the desired peptide (see Fig. 18).

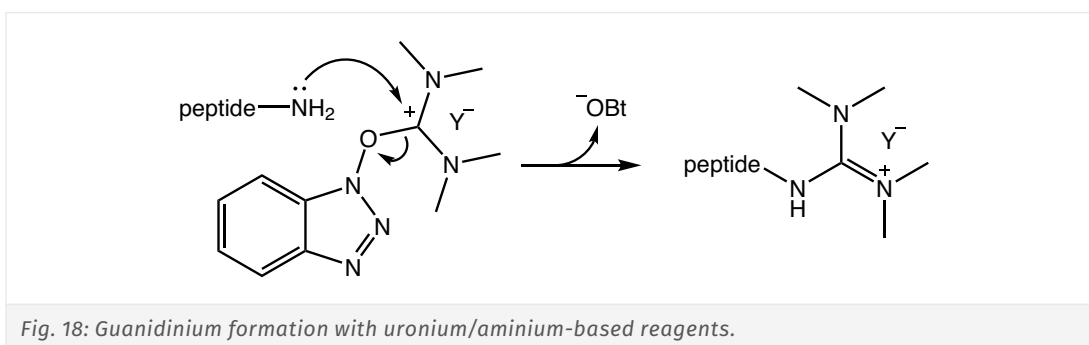


Fig. 18: Guanidinium formation with uronium/aminium-based reagents.

Another consideration is the stability of these reagents upon storage. Uronium/aminium salts are sensitive to moisture and require careful handling as well as storage to maintain their reactivity. In some cases, degradation can lead to lower yields or the need for higher concentrations of reagents.

The HOSu (*N*-hydroxysuccinimide)-based coupling reagents 2-(5-norbornene-2,3-dicarboximido)-1,1,3,3-tetramethyluronium tetrafluoroborate, (TNTU, RL-1063 on page 21) and 2-succinimido-1,1,3,3-tetramethyluronium tetrafluoroborate, (TSTU, RL-1067 on page 22) have been shown to be useful coupling reagents under aqueous reaction conditions.

S-(1-oxido-2-pyridinyl)-1,1,3,3-tetramethylthiuronium hexafluorophosphate, HOTT, [RL-1156 on page 21](#)) and S-(1-oxido-2-pyridinyl)-1,1,3,3-tetramethylthiuronium tetrafluoroborate, (TOTT, [RL-1159 on page 22](#)), derived from 1,1,3,3-tetramethylurea, are used in solution-phase peptide synthesis. These compounds demonstrate comparable results to more expensive coupling reagents in terms of both yield and racemization rates.

2-(2-Oxo-1(2H)-pyridyl)-1,1,3,3-tetramethyluronium tetrafluoroborate (TPTU, [RL-1064 on page 22](#)) is classified as a potentially explosive but not shock-sensitive compound. However, it is also applied in large-scale syntheses, such as the synthesis of an aza-dipeptide analog used in the treatment of AIDS.

Guidelines for uronium/aminium-based coupling procedures

HBTU ([RL-1030 on page 20](#)) / **TBTU** ([RL-1060 on page 21](#)) / **HCTU** ([RL-1031 on page 20](#)) / **HATU** ([RL-1190 on page 20](#))

- I. In a separate flask or tube, dissolve the *N*-protected amino acid (2 eq. relative to the resin loading) in DMF (5 mL/g of resin) and add it to the pre-swelled resin.
- II. Add coupling reagent (2 eq.) and DIPEA (4 eq.) to the solution.
Note: In case of HATU ([RL-1190 on page 20](#)) use 1.9 eq. of coupling reagent but 4 eq. of base.
- III. In order to suppress racemization HOBt or OxymaPure® ([RL-1180 on page 9](#)) (2 eq., 0.5 M solution in DMF) can be added.
- IV. Shake the resin with the coupling mixture for 10-60 min.
- V. Remove the excess reagents after the desired reaction time.
- VI. Wash the resin with DMF (at least 3 x 5 mL).

COMU ([RL-1175 on page 20](#)) (solid phase)

- I. In a separate flask or tube, dissolve the *N*-protected amino acid (3 eq. relative to the resin loading) in DMF (5 mL/g of resin) with COMU ([RL-1175 on page 20](#)) (3 eq., 0.3 M in DMF) and DIPEA (6 eq.).
- II. Pre-activate the mixture for 1 min and add it to the pre-swelled resin.
- III. Shake the resin with the coupling mixture for 10-30 min.
- IV. Remove the excess reagents after the desired reaction time.
- V. Wash the resin with DMF (at least 3 x 5 mL).
- VI. Repeat the procedure in case of sterically-hindered amino acids.

COMU ([RL-1175 on page 20](#)) (solution phase)

- I. Add (0.25 mmol) COMU ([RL-1175 on page 20](#)) to a mixture of *N*-protected amino acid (0.25 mmol), the amino component (0.25 mmol) and base (0.50 mmol or 0.75 mmol in case of ester hydrochloride) in 2 mL DMF at 0 °C.
- II. Stir the reaction mixture for 1 h at 0°C and 2-3 h at room temperature.
- III. After complete reaction, dilute the reaction mixture with EtOAc (25 mL).
- IV. Extract with 1N HCl (2 x 5 mL), 1N NaHCO₃ (2 x 5 mL) and sat. NaCl solution (2 x 5 mL).
- V. Dry the organic phase with MgSO₄ and remove the solvent *in vacuo*.

TSTU ([RL-1067 on page 22](#)) / **HSTU** ([RL-1039 on page 21](#)) / **TNTU** ([RL-1063 on page 21](#)) / **HNTU** ([RL-1155 on page 21](#)) (solution phase)

- I. Dissolve *N*-protected amino acid (1.0 mmol) and of coupling reagent (1.5 mmol) with DIPEA (2 mmol) and amino acid component (2 mmol) in 4 mL DMF.
- II. Stir the reaction for 10 min at room temperature (TLC control).
- III. After complete reaction, dilute the reaction mixture with H₂O (25 mL).
- IV. Extract with DCM (3 x 25 mL).
- V. Dry the organic phase with Na₂SO₄ and remove the solvent *in vacuo*.

[↑ back to content](#)

Introduction

Carbodiimides

Phosphonium Salts

Uronium and Aminium Salts

Acid-Halides

Anhydrides

Triazines

Nyamides

Index

HOTT (RL-1156 on page 21) / TOTT (RL-1159 on page 22) (solution phase)

- I. Dissolve N-protected amino acid (1 mmol) and the amino component (1 mmol) in 5 mL DMF.
- II. Add DIPEA (2 mmol) and coupling reagent (1 mmol) to the mixture.
- III. Stir the reaction for 30 min at room temperature (TLC control).
- IV. After complete reaction, dilute the reaction mixture with sat. NaCl (25 mL).
- V. Extract with EtOAc (3 x 25 mL).
- VI. Wash the combined organic phases with 2N HCl (2 x 10 mL), sat. NaHCO₃ (2 x 10 mL) and H₂O (6 x 10 mL).
- VII. Dry the organic phase with Na₂SO₄ and remove the solvent *in vacuo*.

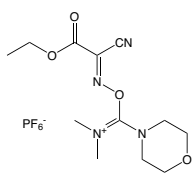

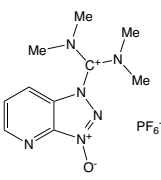

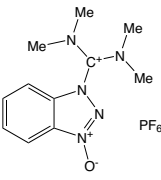

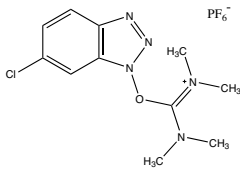

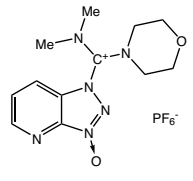

TPTU (RL-1064 on page 22) (solution phase)

- I. Dissolve N-protected amino acid (3 eq.) and TPTU (RL-1064 on page 22) (3 eq.) in DCM (3 mL, overall concentration 1 M).
- II. Cool the resulting solution to 0°C.
- III. Add DIPEA (6 eq.) to the mixture.
- IV. Add the amino component (3 eq.) to the generated active ester portion wise at 0-5 °C.
- V. Remove the ice bath and let the reaction mixture stir at room temperature overnight.
- VI. Wash the reaction mixture with H₂O (10mL), sat. NaHCO₃ (10 mL), and sat. NaCl (5 mL).
- VII. Extract the aqueous phases with DCM (2 x 5 mL).
- VIII. Dry the organic phase with Na₂SO₄ and remove the solvent *in vacuo*.


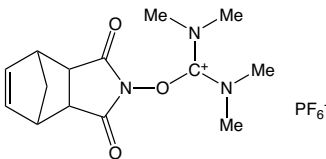

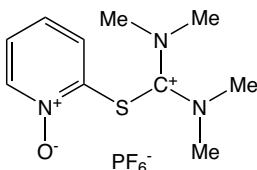

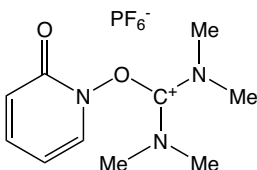

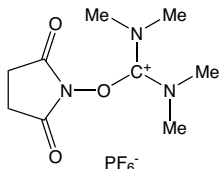

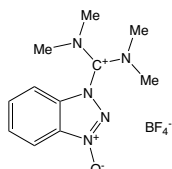

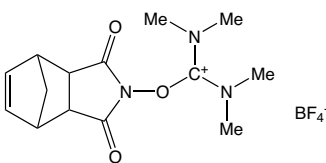
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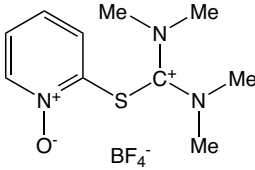

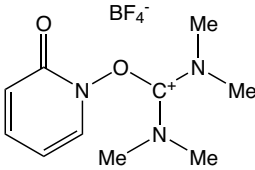

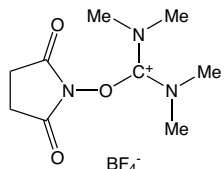

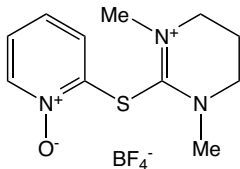

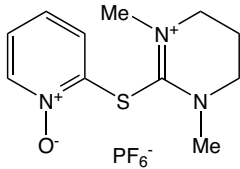

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		Product details	
RL-1175	COMU®		
1-[1-(Cyano-2-ethoxy-2-oxoethylideneaminoxy)-dimethylamino-morpholino]-uronium hexafluorophosphate			
CAS-No.	1075198-30-9		
Formula	C ₁₂ H ₁₉ N ₄ O ₄ PF ₆		
Mol. weight	428,26 g/mol		
RL-1190	HATU		
2-(1H-7-Azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate			
CAS-No.	148893-10-1		
Formula	C ₁₀ H ₁₅ N ₆ O*PF ₆		
Mol. weight	235,27*144,96 g/mol		
RL-1030	HBTU		
2-(1H-Benzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate			
CAS-No.	94790-37-1		
Formula	C ₁₁ H ₁₆ N ₅ O*PF ₆		
Mol. weight	234,28*144,96 g/mol		
RL-1031	HCTU		
O-(1H-6-Chlorobenzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate			
CAS-No.	330645-87-9		
Formula	C ₁₁ H ₁₅ ClF ₆ N ₃ OP		
Mol. weight	413,69 g/mol		
RL-2170	HDMA		
1-[(Dimethylamino)(morpholino)methylene]-1H-[1,2,3] triazolo[4,5-b]pyridine-1-ium 3-oxide hexafluorophosphate			
CAS-No.	958029-37-3		
Formula	C ₁₂ H ₁₇ N ₆ O ₂ *PF ₆		
Mol. weight	277,30*144,96 g/mol		

[↑ back to content](#)

		Product details
RL-1155	HNTU	
2-(endo-5-norbornene-2,3-dicarboxymido)-1,1,3,3-tetramethyluronium hexafluorophosphate		
CAS-No.	208462-94-6	
Formula	$C_{14}H_{20}N_3O_3 \cdot PF_6^-$	
Mol. weight	278,33*144,96 g/mol	
		
RL-1156	HOTT	
S-(1-Oxido-2-pyridyl)-thio-N,N,N',N'-tetramethyluronium hexafluorophosphate		
CAS-No.	212333-72-7	
Formula	$C_{10}H_{16}N_3OS \cdot PF_6^-$	
Mol. weight	226,32*144,96 g/mol	
		
RL-1092	HPTU	
O-(1,2-Dihydro-2-oxo-pyridyl)-N,N,N',N'-tetramethyluronium hexafluorophosphate		
CAS-No.	364047-51-8	
Formula	$C_{10}H_{16}N_3O_2 \cdot PF_6^-$	
Mol. weight	210,26*144,96 g/mol	
		
RL-1039	HSTU	
2-Succinimido-1,1,3,3-tetramethyluronium hexafluorophosphate		
CAS-No.	265651-18-1	
Formula	$C_9H_{16}N_3O_3 \cdot PF_6^-$	
Mol. weight	359,21 g/mol	
		
RL-1060	TBTU	
2-(1H-Benzotriazole-1-yl)-1,1,3,3-tetramethyluronium tetrafluoroborate		
CAS-No.	125700-67-6	
Formula	$C_{11}H_{16}N_5O \cdot BF_4^-$	
Mol. weight	234,28*86,81 g/mol	
		
RL-1063	TNTU	
2-(endo-5-norbornene-2,3-dicarboxymido)-1,1,3,3-tetramethyluronium tetrafluoroborate		
CAS-No.	125700-73-4	
Formula	$C_{14}H_{20}N_3O_3 \cdot BF_4^-$	
Mol. weight	278,33*86,80 g/mol	
		

			Product details
RL-1159	TOTT		
S-(1-Oxo-2-pyridyl)-thio-N,N,N',N'-tetramethyluronium tetrafluoroborate			
CAS-No.	255825-38-8		
Formula	C ₁₀ H ₁₆ N ₃ OS*BF ₄		
Mol. weight	226,32*86,80 g/mol		
			
RL-1064	TPTU		
O-(1,2-Dihydro-2-oxo-pyridyl)-N,N,N',N'-tetramethyluronium tetrafluoroborate			
CAS-No.	125700-71-2		
Formula	C ₁₀ H ₁₆ N ₃ O ₂ *BF ₄		
Mol. weight	210,25*86,80 g/mol		
			
RL-1067	TSTU		
N,N,N',N'-Tetramethyl-O-(succinimidyl)uronium tetrafluoroborate			
CAS-No.	105832-38-0		
Formula	C ₉ H ₁₆ N ₃ O ₃ *BF ₄		
Mol. weight	214,24*86,80 g/mol		
			
RL-1161	TPTDP		
S-(1-Oxo-2-pyridyl)-thio-1,3-dimethylpropyleneuronium tetrafluoroborate			
CAS-No.	367252-09-3		
Formula	C ₁₁ H ₁₆ N ₃ OS*BF ₄		
Mol. weight	238,33*86,80 g/mol		
			
RL-1158	HPTDP		
S-(1-Oxo-2-pyridyl)-thio-1,3-dimethylpropyleneuronium hexafluorophosphate			
CAS-No.	366821-62-7		
Formula	C ₁₁ H ₁₆ N ₃ OS*PF ₆		
Mol. weight	238,33*144,96 g/mol		
			

Introduction

Carbodimides

Phosphonium Salts

Uronium and Aminium Salts

Acid-Halides

Anhydrides

Triazines

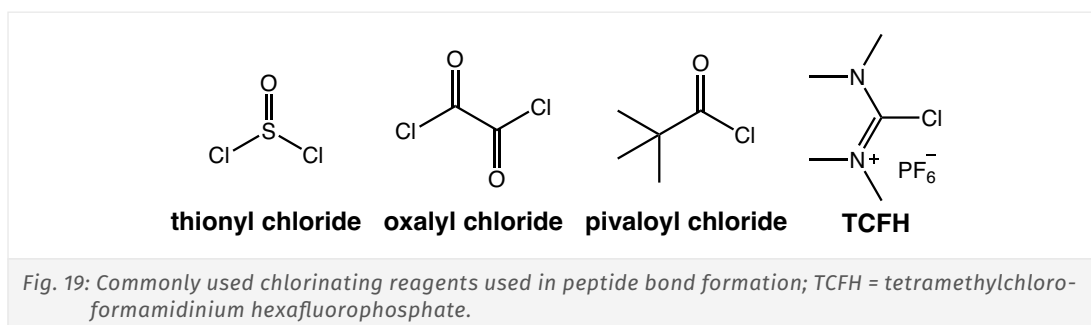
Vnamides

Index

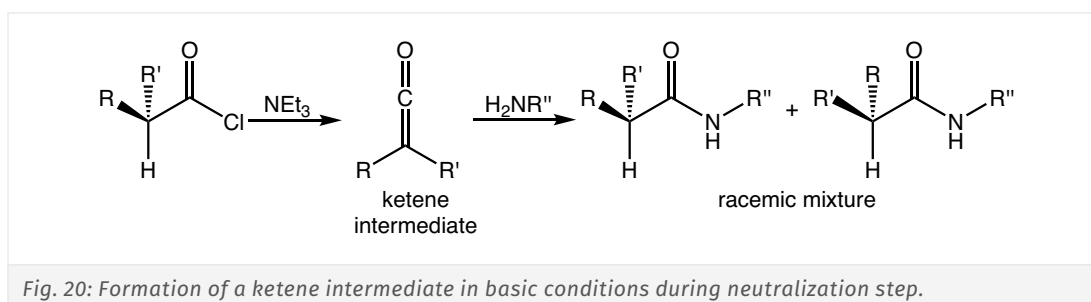
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5. Acid-Halides

The activation of carboxylic acids *via* carboxylic halides, especially acid chlorides, followed by their reaction with free amines, is one of the most well-established methods for peptide bond formation. The high reactivity of acid chlorides enables rapid and efficient coupling reactions, making them especially useful when working with bulky or less nucleophilic amino acids. However, this reactivity often comes at the expense of chiral integrity, as epimerization might occur under the coupling conditions. In large-scale peptide synthesis, reagents such as thionyl chloride (SOCl_2), oxalyl chloride ($(\text{COCl})_2$) and pivaloyl chloride are commonly applied for activation due to their high effectiveness (see Fig. 19).



Among these, thionyl chloride, particularly when used in pyridine, is one of the most widely utilized chlorinating agents. In the context of Fmoc-protected amino acids, thionyl chloride is typically used in heated dichloromethane (DCM). A notable disadvantage of this approach is the generation of hydrogen chloride which is crucial when using acid-labile protecting groups. This by-product necessitates neutralization with a base, a step that can inadvertently facilitate the formation of oxazolones (see Fig. 2) or ketenes (see Fig. 20), ultimately resulting in racemization.

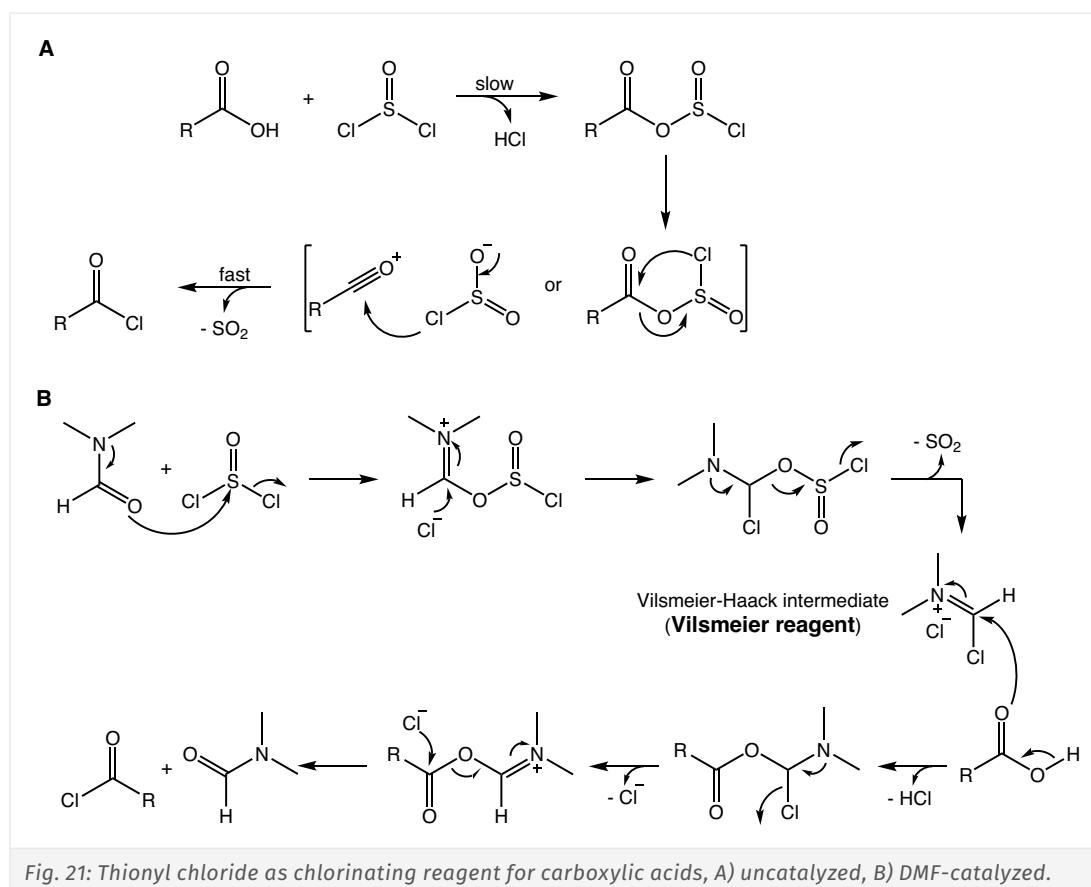


The addition of a small amount of DMF during the formation of acid chlorides with thionyl chloride can serve as a catalyst, forming a Vilsmeier-Haack intermediate (commonly referred to as the Vilsmeier reagent) (see Fig. 21).

After activation, the peptide bond formation step is performed under inert conditions.

Note:

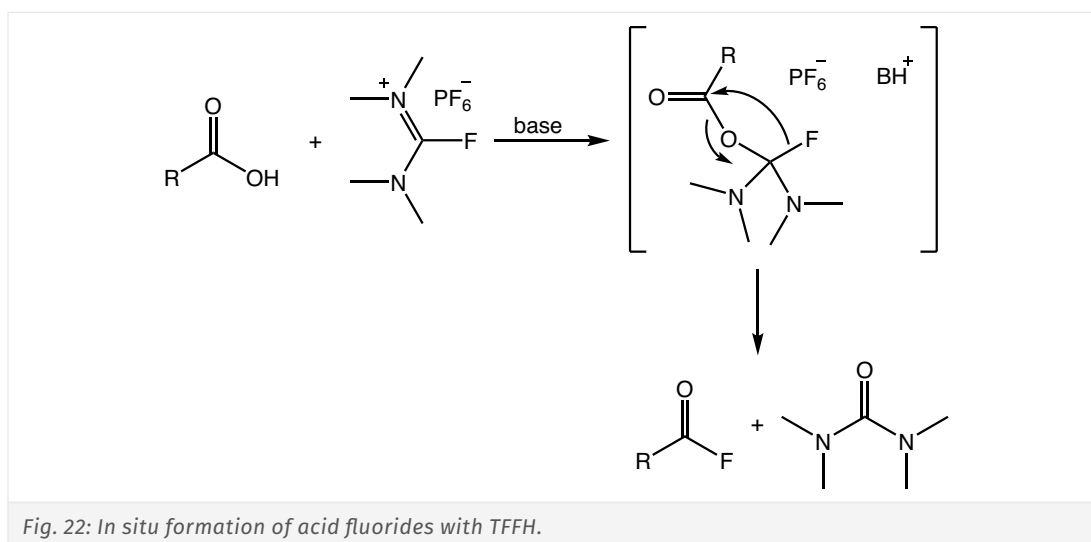
Despite its widespread use due to cost-effectiveness, thionyl chloride poses the risk of producing dimethylcarbamoyl chloride, a potentially carcinogenic by-product.



Chlor-*N,N,N',N'*-tetramethylformamidium-hexafluorophosphat (TCFH, [RL-1071 on page 27](#)) is another chlorinating reagent used as precursor for the synthesis of other coupling reagents such as HATU ([RL-1190 on page 20](#)) or HCTU ([RL-1031 on page 20](#)). However, it can also be used directly as a coupling reagent in ACN or water, for example, but also in the synthesis on solid support.

Acid fluorides exhibit slower rates in oxazolone formation in the presence of tertiary amines and provide greater stability against moisture and acid-labile functional groups compared to their chloride counterparts. Fluoroformamidinium salts, such as tetramethylfluoroformamidinium hexafluorophosphate (TFFH, [RL-1077 on page 27](#)), are highly effective for *in situ* acid fluoride generation.

TFFH's air-stable, non-hygroscopic nature makes it an ideal reagent for peptide synthesis. This method is especially beneficial for amino acids like arginine (Arg) and histidine (His), as their acid fluorides are too unstable for direct isolation or extended storage. Recently, TFFH ([RL-1077 on page 27](#)) was declared as a green coupling reagent.



Guidelines for coupling procedures

Acyl halides

Coupling reactions are typically conducted in inert dry solvents alongside non-nucleophilic tertiary amines such as NEt_3 , DIPEA or NMM. Common organic solvents employed in these reactions include THF, n-heptane, toluene, ACN, or dimethoxyethane (DME). Notably, acyl chlorides exhibit sufficient stability to undergo coupling with amines under aqueous conditions, by utilizing bases like NaOH , NaHCO_3 , K_2CO_3 under Schotten–Baumann conditions. The reaction kinetics can be further enhanced by the addition of catalytic amounts of pyridine or *N,N*-dimethylaminopyridine (DMAP), with pyridine sometimes used as the solvent.

General procedure

- I. In an inert atmosphere dissolve the acyl chloride (2 mmol) in dry THF (4 mL).
- II. Cool the solution to 0 °C.
- III. Add potassium phosphate (5 mmol) at once and add the amino acid (2 mmol) to the reaction mixture.
- IV. Stir the resulting mixture for 12 h at room temperature.
- V. The reaction was quenched with water (10 mL) and EtOAc (4 mL).
- VI. The organic phase can be discarded, while the pH of the aqueous phase is adjusted to pH = 2.
- VII. Extract with EtOAc (2 x 10 mL) and wash with water (6 mL).
- VIII. Dry the organic phase with MgSO_4 and remove the solvent *in vacuo*.

TCFH (solution phase) ([RL-1071 on page 27](#))

- I. Dissolve the *N*-protected amino acid (1 eq.), the amino acid component (1.3 eq.), and *N*-methylimidazole (3.5 eq.) in ACN (4 mL).
- II. Add TCFH (1.2 eq.) in a single portion.
- III. Stir the resulting mixture for 19 h at room temperature.
- IV. After completion of the reaction, add isopropyl acetate (6 mL) and H_2O (4 mL) and separate the phases.
- V. Extract the aqueous phase with isopropyl acetate (4 mL).
- VI. Wash the combined organic phases with water (6 mL).
- VII. Dry the organic phase with MgSO_4 and remove the solvent *in vacuo*.

TFFH ([RL-1077 on page 27](#))

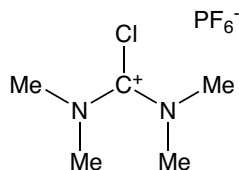

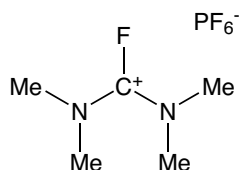

- I. In a separate flask or tube, dissolve the N-protected amino acid (5 eq. relative to the resin loading) in DMF.
- II. Add TFFH ([RL-1077 on page 27](#)) (5 eq.) and DIPEA (10 eq.) to the solution (total concentration 0.3 M).
- III. Pre-activate the resulting mixture for 8-15 min at room temperature. Note: Complete conversion of hindered amino acids such as Aib might take 1-2 h.
- IV. Add it to the pre-swelled resin and shake the resin with the coupling mixture for 30 min. Note: For sterically-hindered amino acids (e.g., Aib) use a coupling time of 60 min or repeat the procedure.
- V. Remove the excess reagents after the desired reaction time.
- VI. Wash the resin with DMF (at least 3 x 5 mL).

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[↑ back to content](#)

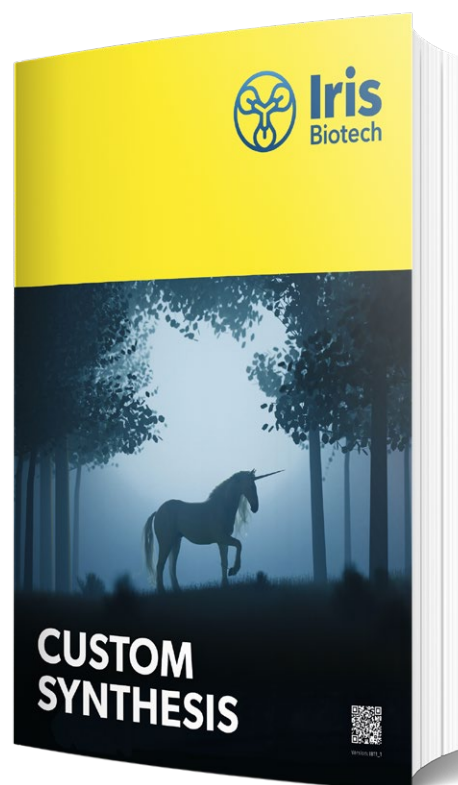
Coupling Reagents

			Product details
RL-1071	TCFH		
N,N,N',N'-Tetramethylchloroformamidinium-hexafluorophosphate			
CAS-No.	207915-99-9		
Formula	C ₅ H ₁₂ ClN ₂ *PF ₆		
Mol. weight	135,61*144,96 g/mol		
RL-1077	TFFH		
N,N,N',N'-Tetramethylfluoroformamidinium hexafluorophosphate			
CAS-No.	164298-23-1		
Formula	C ₅ H ₁₂ FN ₂ *PF ₆		
Mol. weight	119,16*144,96 g/mol		



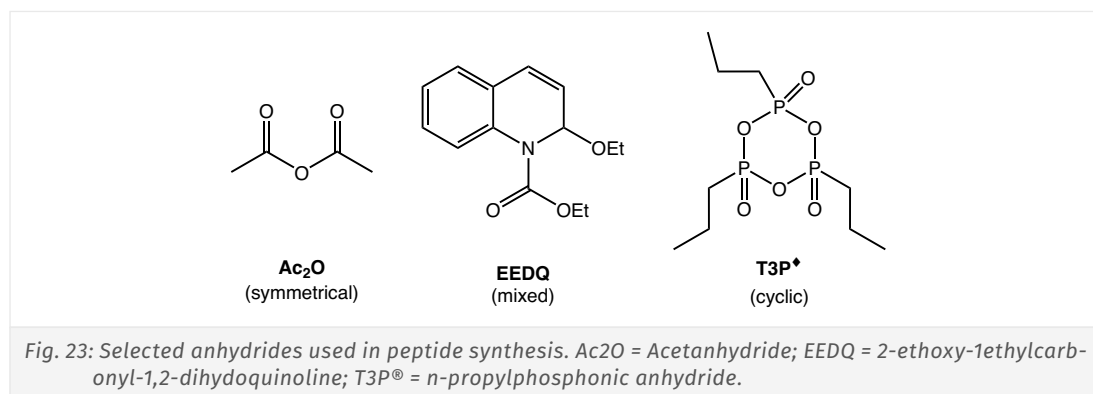
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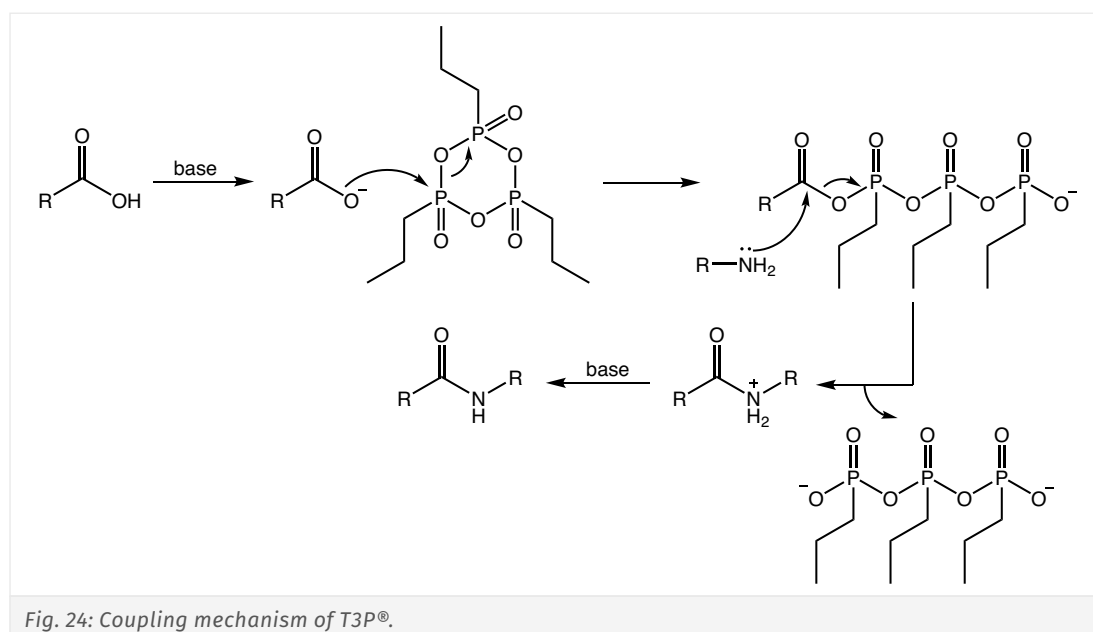
6. Anhydrides

Peptide bond formation by anhydrides is one of the oldest and most established methods in peptide chemistry. Anhydrides can be classified into symmetrical, mixed, and cyclic types (see Fig. 23), each offering distinct properties and applications in peptide synthesis.



In recent years, n-propylphosphonic anhydride (T3P®) has become increasingly popular, especially for large/industrial-scale processes peptide synthesis. Like TFFH ([RL-1077 on page 27](#)) (chapter 5), T3P® is recognized as an environmentally friendly (“green”) coupling reagent. It is widely used in solution phase synthesis for amide bond formation. T3P® offers several notable advantages, including low toxicity, long shelf-life, and an easy handling.

A key advantage of T3P® is that it produces water-soluble side products, which can be easily removed by extraction into the aqueous phase, greatly simplifying purification (see Fig. 24). Additionally, T3P® induces minimal epimerization, making it ideal for synthesizing peptides with sensitive α-stereocenters



[↑ back to content](#)

Guidelines for coupling with T3P®

- I. In a separate flask or tube, dissolve the *N*-protected amino acid (5 eq. relative to the resin loading) in DMF (1 mL/100 mg of resin) and sonicate for 10 min.
- II. Add DIPEA (10 eq.) and add T3P® (5 eq.).
- III. Add the resulting mixture to the pre-swelled resin.
- IV. Add OxymaPure® (5 eq.).
- V. Shake the resin with the coupling mixture for 60 min at 60 °C.
- VI. Remove the excess reagents after the desired reaction time.
- VII. Wash the resin with DMF (at least 3 x 5 mL).
- VIII. Repeat the procedure in case of sterically-hindered amino acids.

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7. Triazines

Triazines, for example derivatives such as 2,4,6-trichloro-1,3,5-triazine (cyanuric chloride) and 2-chloro-4,6-dimethoxy-1,3,5-triazine (CDMT), are widely utilized as efficient coupling reagents in peptide bond formation. These heterocyclic compounds (see [Fig. 25](#)), consisting of a six-membered ring structure with alternating carbon and nitrogen atoms, serve to activate carboxyl groups, facilitating amide bond formation by promoting the nucleophilic attack by amines.

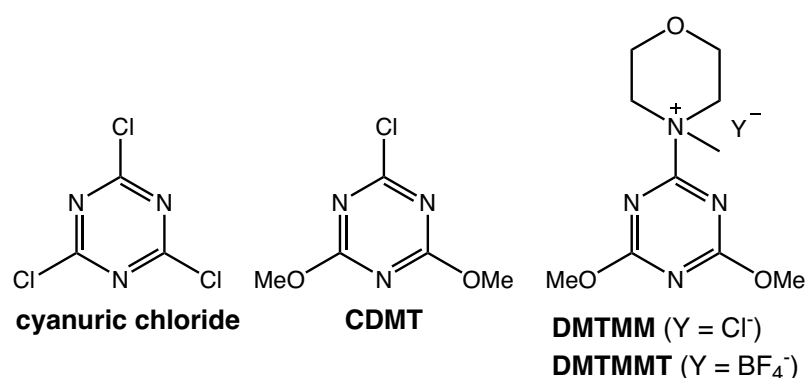
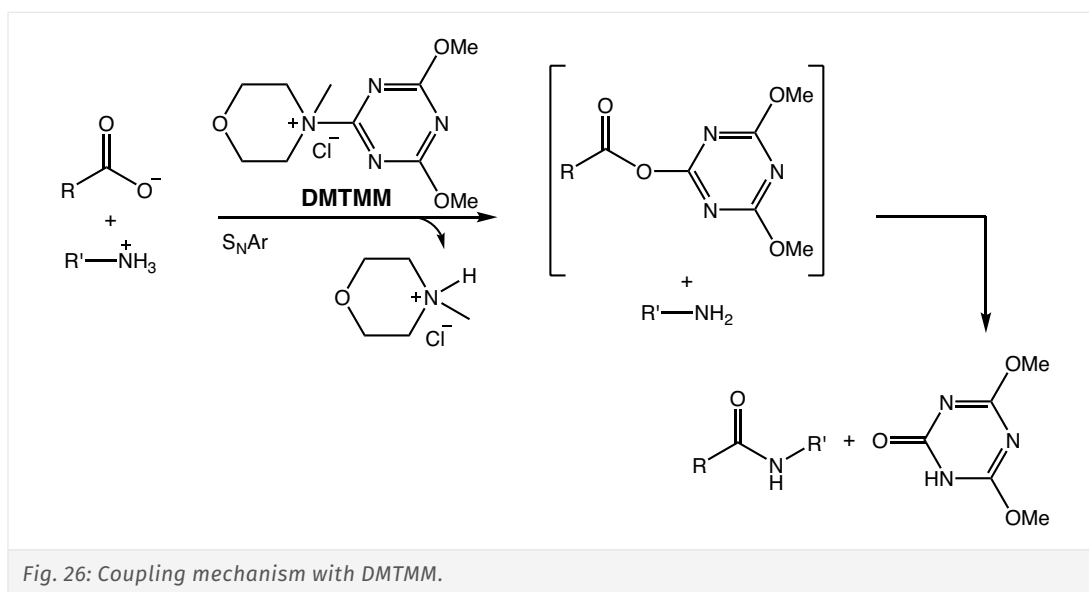


Fig. 25: Chosen triazine coupling reagents.

Cyanuric chloride plays a key role in the synthesis of acyl chlorides, amides, and peptides, acting as a versatile platform for derivatization. Its derivative, CDMT is known for forming highly reactive esters with carboxylic acids, making it an effective acylating agent for amines and, to a lesser extent, alcohols. This activation is typically achieved in the presence of a base, such as NMM, which enhances the reactivity of CDMT. The reaction between NMM and CDMT produces the intermediate 4-(4,6-dimethoxy-1,3,5-triazin-2-yl)-4-methylmorpholin-1-ium chloride (DMTMM), which can be isolated and utilized independently. This feature eliminates the need for prior activation of carboxylic acids, simplifying the synthesis process.

DMTMM has emerged as an efficient coupling agent in peptide synthesis, offering high product yields in one-step reactions under mild conditions (room temperature and atmospheric pressure). Notably, it exhibits compatibility with a range of easily removable solvents such as methanol, ethanol, isopropanol, and water, eliminating the need for rigorous solvent drying procedures. The reaction proceeds without requiring additional additives, and carboxylic acids can be activated *in situ*, broadening its utility in various synthetic applications. Furthermore, DMTMM has demonstrated comparable coupling efficiency to commonly used peptide synthesis reagents like PyBOP ([RL-2005 on page 15](#)), while minimizing racemization to undetectable levels during solid-phase peptide synthesis.

The reaction mechanism of DMTMM is based on nucleophilic aromatic substitution (S_NAr). In this mechanism, the activated ester intermediate is displaced by the amine, leading to amide bond formation (see [Fig. 26](#)). NMM is simultaneously regenerated as a by-product, eliminating the need for an additional base in the reaction. The secondary by-product, triazinone, can be easily removed by aqueous washing, simplifying post-reaction purification.



However, a notable limitation of this reaction is the release of one equivalent of NMM.

Various studies have explored alternative tertiary amines to replace NMM, evaluating their reactivity alongside DMTMM-like reagents.

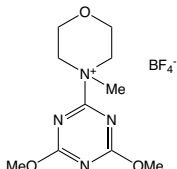

Guidelines for coupling with DMTMMT

- I. In a separate flask or tube, dissolve the *N*-protected amino acid (3 eq. relative to the resin loading) in DMF (5 mL/g of resin).
- II. Add DMTMMT ([RL-1105 on page 32](#)) (3 eq.) and NMM (6 eq.).
- III. Add the resulting mixture to the pre-swelled resin.
- IV. Shake the resin with the coupling mixture for 15 min (perform Kaiser-test).
- V. Remove the excess reagents after the desired reaction time.
- VI. Wash the resin with DMF and DCM (2 x 5 mL for 2 min).

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- Synthesis of sterically-hindered peptidomimetics using 4-(4,6-dimethoxy-1,3,5-triazine-2-yl)-4-methyl-morpholinium chloride; W.-C. Shieh, Z. Chen, S. Xue, J. McKenna, R.-M. Wang, K. Prasad, O. Repič; *Tetrahedron Letters* 2008; **49**(37): 5359-5362. <https://doi.org/10.1016/j.tetlet.2008.06.119>

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		Product details
RL-1105	DMTMMT	
4-(4,6-Dimethoxy-1,3,5-triazin-2-yl)-4-methyl-morpholinium tetrafluoroborate		
CAS-No.	293311-03-2	
Formula	C ₁₀ H ₁₇ BF ₄ N ₄ O ₃	
Mol. weight	241,27*86,80 g/mol	
		


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8. Ynamides

Ynamides are novel coupling reagents characterized by a triple bond to a nitrogen atom that is substituted with an electron-withdrawing group (see Fig. 27).

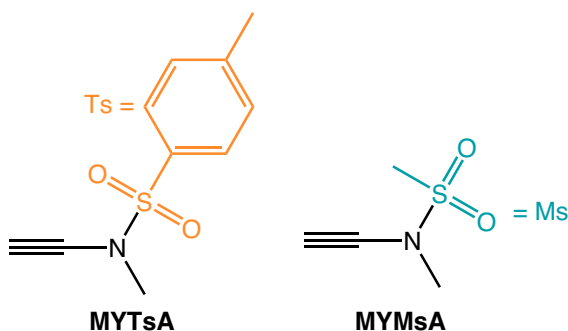


Fig. 27: Prominent ynamide coupling reagents.

These compounds can be utilized in a two-step, one-pot methodology to facilitate the formation of peptide bonds. This process involves the efficient hydroacylation of ynamides with carboxylic acids, followed by aminolysis (see Fig. 28). Notably, these reactions occur under mild conditions, allowing for the exclusion of a base, which prevents the deprotonation of the C α atom and avoids the formation of oxazolones, thereby eliminating the risk of racemization.

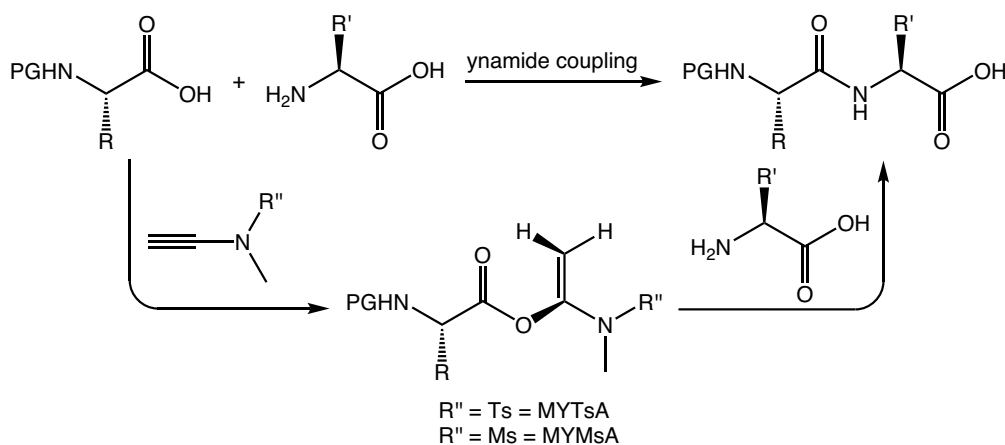
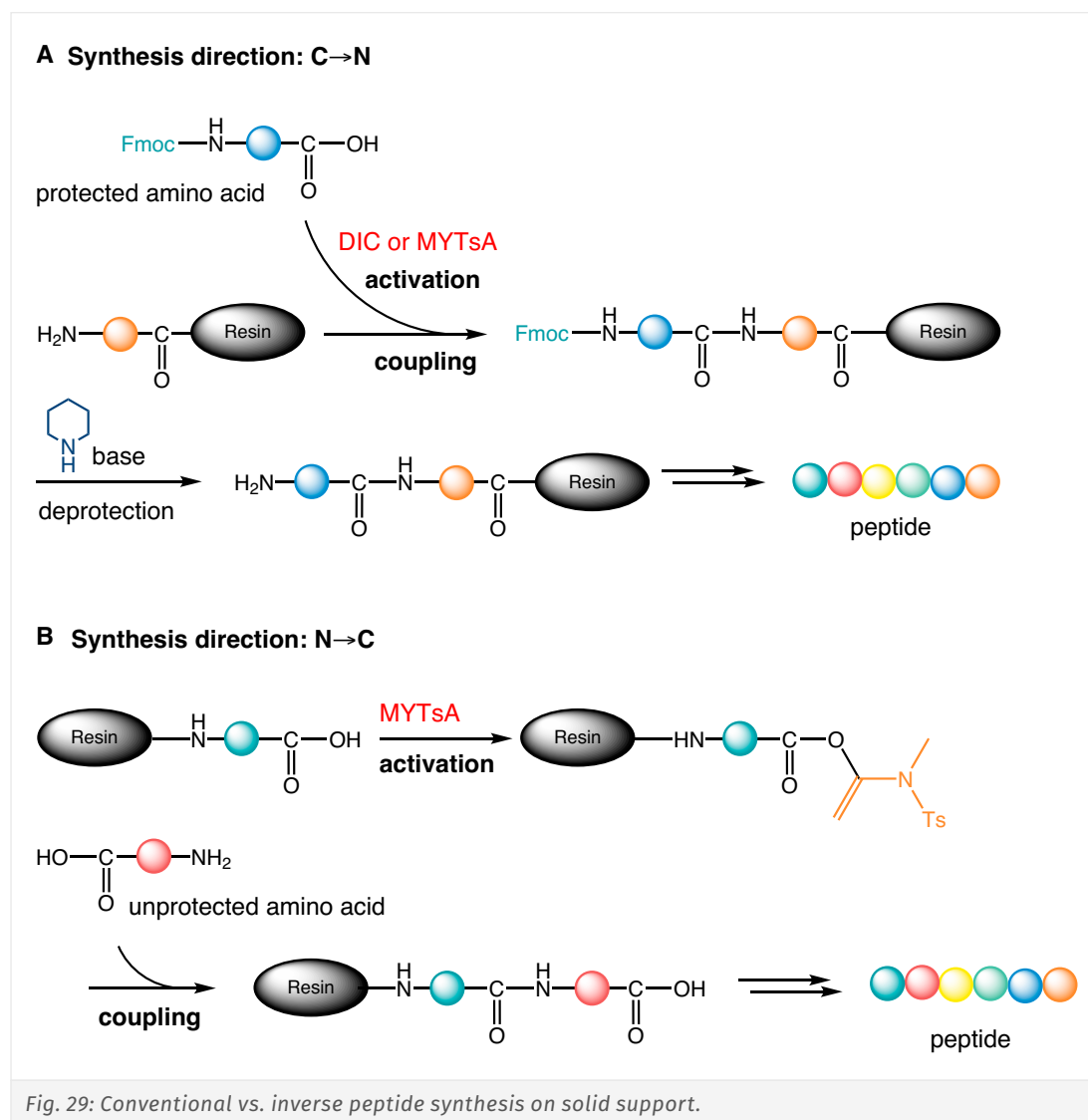


Fig. 28: Two-step, one-pot procedure of ynamide coupling.

Furthermore, ynamides exhibit stability under both air and moisture and can be applied without the need for additives or catalysts, rendering them suitable for peptide segment condensations and large-scale synthesis. Additionally, ynamides demonstrate compatibility with various functional groups found in side-chains of amino acids, including hydroxyl groups (serine, threonine), thiols (cysteine), amides (asparagine, glutamine), and the amine functionality of indole (tryptophane).

MYTsA (RL-8665 on page 35) offers an additional advantage by enabling peptide synthesis in the N → C direction, mirroring natural protein biosynthesis (see Fig. 29). This method avoids the issues of racemization associated with carbodiimides and offers a superior atom economy, resulting in less waste. Furthermore, the amino groups of the amino acid building blocks do not require protection or deprotection in this process, thereby eliminating the need for Fmoc protecting group and base (commonly piperidine).


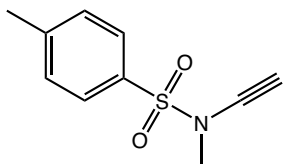


Coupling guidelines

- I. Dissolve MYTsA (0.2 mmol) and the *N*-protected amino acid (0.2 mmol) in DCM (1 mL).
- II. Stir the resulting mixture until full consumption of MYTsA (TLC control).
- III. Add the amino acid (0.2 mmol) to the formed α -acyloxyenamide.
- IV. Stir the reaction mixture until full consumption of α -acyloxyenamide (TLC control).
- V. Concentrate the reaction mixture *in vacuo* and purify the crude product by silica gel chromatography.

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		Product details
RL-8665	MYTsA	
N-ethynyl-N,4-dimethylbenzenesulfonamide		
CAS-No.	1005500-75-3	
Formula	C ₁₀ H ₁₁ NO ₂ S	
Mol. weight	209,26 g/mol	
		

Index

Product code	Product name	Page
RL-5096	BOP	14
RL-1175	COMU ®	20
RL-1013	DCC	9
RL-1153	DEPBT	14
RL-1015	DIC	9
RL-1105	DMTMMT	32
RL-1022	EDC*HCl	9
RL-1190	HATU	20
RL-1030	HBTU	20
RL-1031	HCTU	20
RL-2170	HDMA	20
RL-1155	HNTU	21
RL-1091	HOCl	10
RL-1036	HONB	10
RL-2002	HOPO	10
RL-1156	HOTT	21
RL-1158	HPTDP	22
RL-1092	HPTU	21
RL-1039	HSTU	21
RL-8665	MYTsA	35
RL-2290	Oxyma-B	9
RL-1180	OxymaPure	9
RL-1210	PyAOP	15
RL-2005	PyBOP	15
RL-1051	PyBrOP	15
RL-2180	PyCLOCK	15
RL-2270	PyOxim	14
RL-3910	T-BEC	9
RL-1060	TBTU	21
RL-1071	TCFH	27
RL-1077	TFFH	27
RL-1063	TNTU	21
RL-1159	TOTT	22
RL-1161	TPTDP	22
RL-1064	TPTU	22
RL-1067	TSTU	22

Introduction

Carbodiimides

Phosphonium Salts

Uronium and Aminium Salts

Acid-Halides

Anhydrides

Triazines

Ynamides

Index

[↑ back to content](#)

Notes

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In 2001 Iris Biotech GmbH was founded just at the beginning of the Biotech movement and the first remarkable breakthrough of biotech pharma products. Although the biotech field is rather young compared to other industries we believe on long-term business, a good partnership between our business partners and Iris Biotech GmbH and a good reputation. It is our duty as well as our responsibility to maintain and to extend this over the next generations – based on the principles of an honourable and prudent tradesman which based upon the concept of honourable entrepreneurship.

This Code of Conduct has been developed following the “Voluntary Guidelines for Manufacturers of Fine Chemical Intermediates and Active Ingredients” issued by AIME (Agrochemical & Intermediates Manufacturers in Europe) and the requirements of some of our business associates.

Iris Biotech GmbH commits to hold this Code of Conduct and to include and apply its principles in the management system and the company policies.

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Peptide Solutions LLC

India, Bangladesh, Oman, Sri Lanka, United Arab Emirates:

Sumit Biosciences Pvt Ltd.

Empowering Peptide Innovation